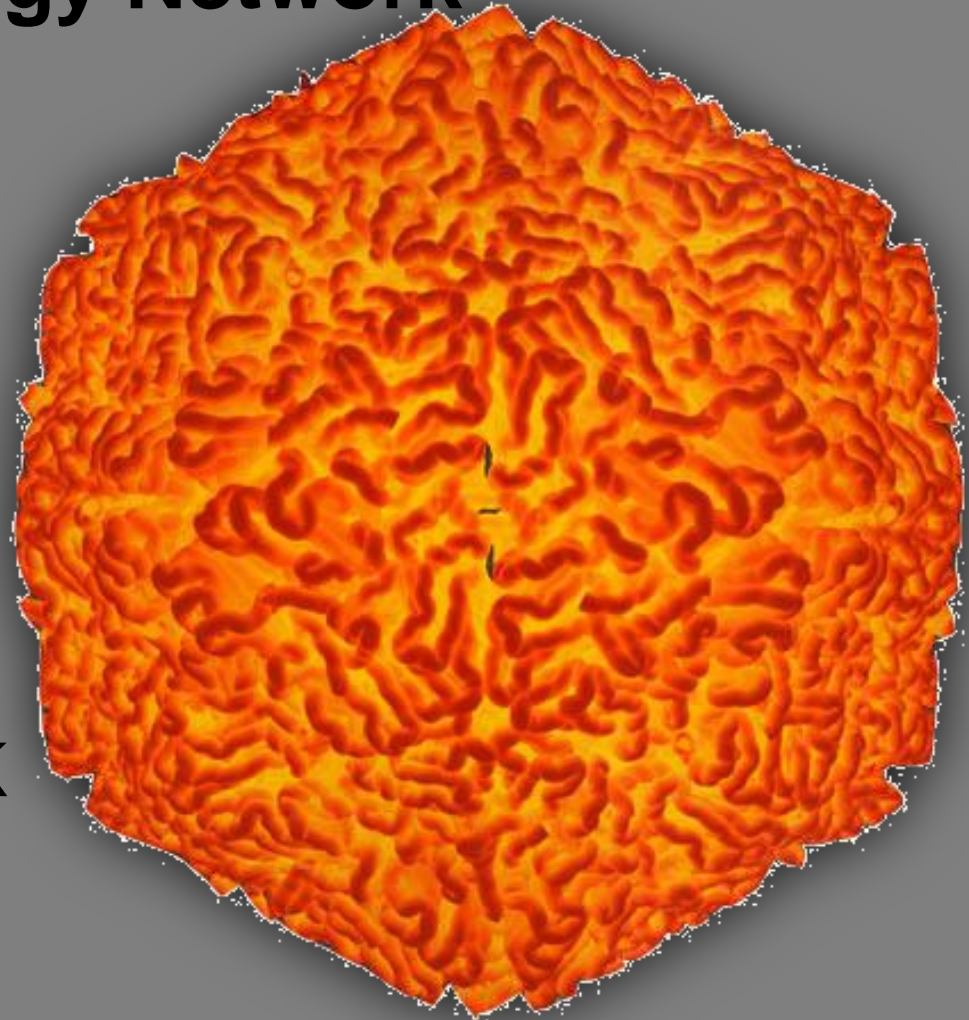


# Structural Biology and Vaccinology

## Veterinary Vaccinology Network

16<sup>th</sup>-17<sup>th</sup> February 2015

DAVID STUART  
Diamond Light Source, UK  
Oxford University, UK  
Instruct, EU



# **Structural vaccinology: a three-dimensional view for vaccine development.**

[Cozzi R<sup>1</sup>, Scarselli M, Ferlenghi I.](#)

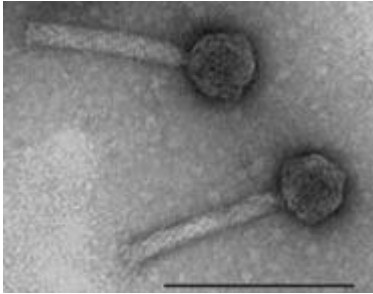
<sup>1</sup>Novartis Vaccines and Diagnostics, Siena, Italy.  
ilaria.ferlenghi@novartis.com

The Structural Vaccinology approach is the logical evolution of Reverse Vaccinology: a genome-based approach combined with structural biology, with the idea that protective determinants can be used to selectively engineer the antigens that can be re-designed and simplified for inclusion in vaccine combinations. The final objectives of the rational structure-based antigen optimization are the facilitation of industrial-scale production of the antigens combination, obtain a greater immunogenicity and a greater safety profile and finally expand the breadth of protection. Structural Vaccinology is particularly powerful in case of antigenic variation between closely related strains and species.

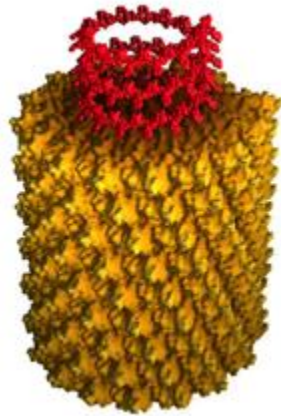
I will only consider viruses ...

# But viruses span a huge range of sizes and shapes

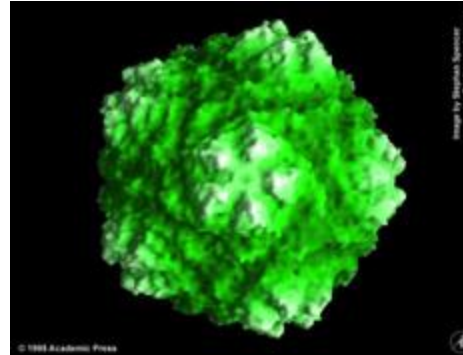
...



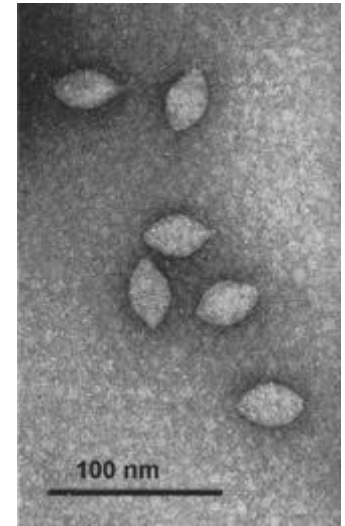
Enterobacteria virus P2



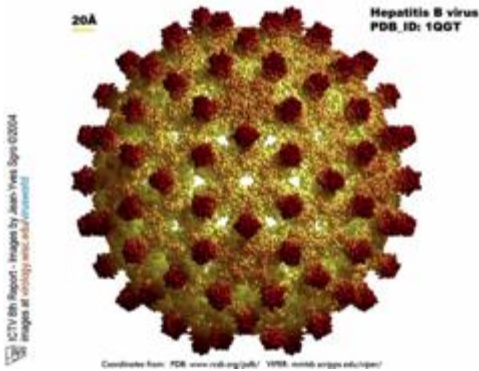
Tobacco mosaic virus



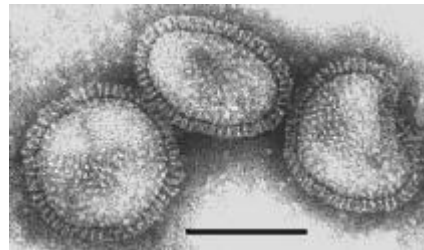
Satellite tobacco necrosis virus



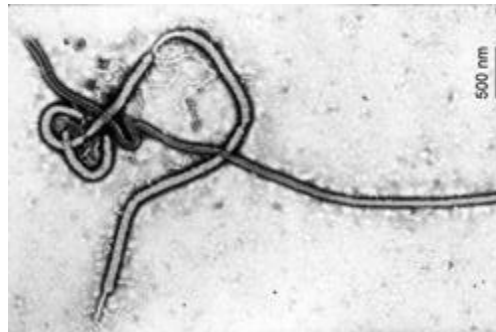
*Sulfolobus* spindle-shaped virus 1



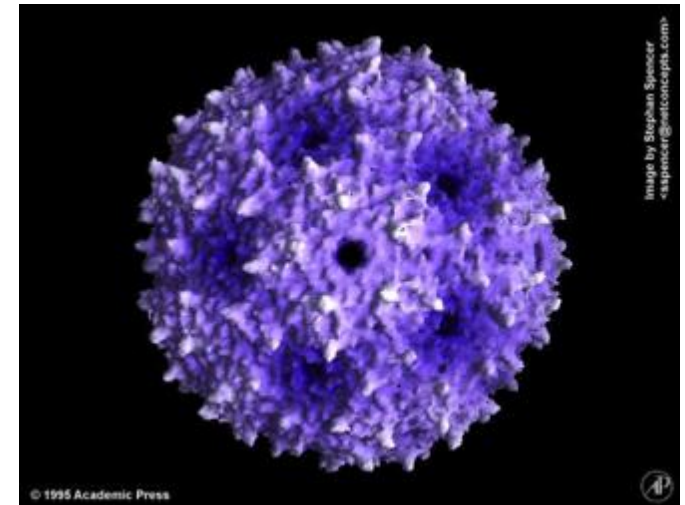
Hepatitis B virus



Influenza virus A



Ebola virus



Enterobacteria phage MS2

Some  $10^{24}$  viral infections are thought to occur every second in the biosphere, a snapshot of a process likely to have been ongoing for several billion years... so the diversity is hardly surprising!

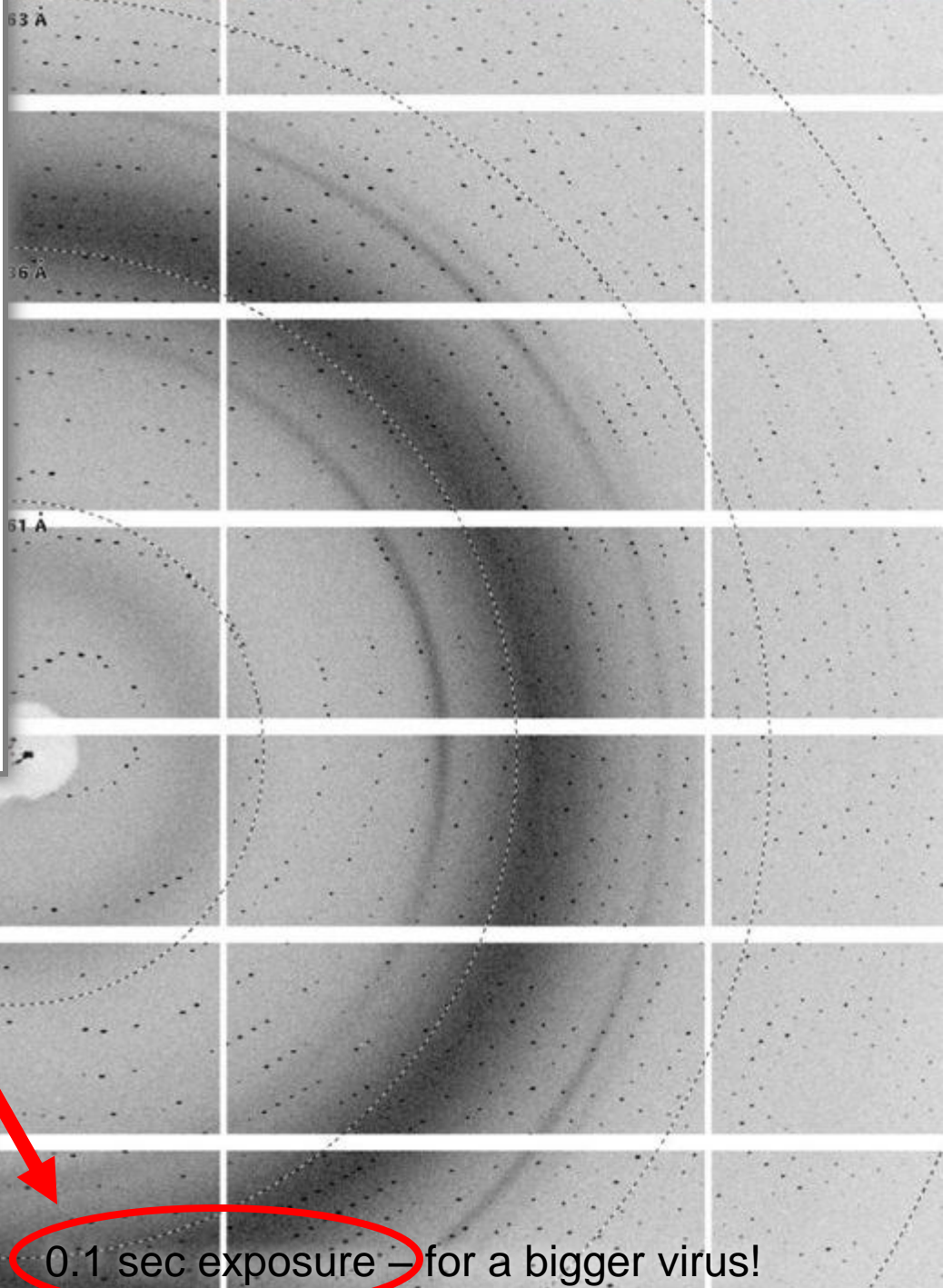
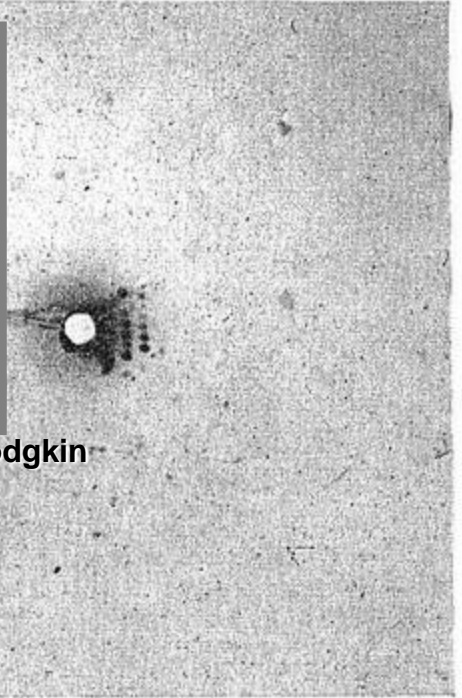
I will talk about work on only a minute fraction of these – picornaviruses – there is no silver bullet – each case will be a bit different

Pharma is used to structure based drug design, but vaccines?

We can now determine the atomic structure of COMPLETE viruses relatively easily, and the question is does this allow us to design new/better vaccines?

The technology advances to enable this are remarkable ...





Dorothy Crowfoot Hodgkin

Fig. 1. TOBACCO NECROSIS VIRUS DERIVATIVE.  
2½° oscillation photograph, crystal rotating about normal to (001), X-ray beam parallel to (100) 90 hours exposure.

1945

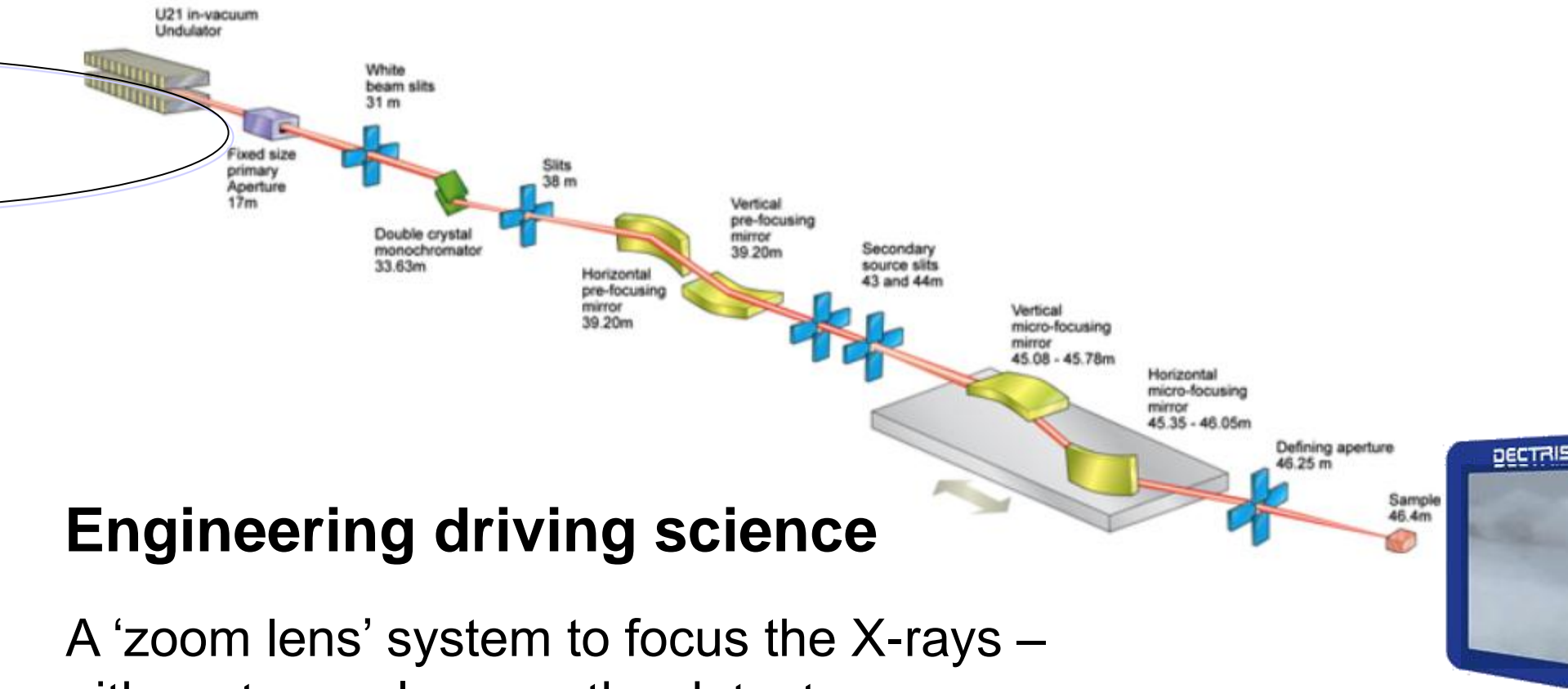
2012



Diamond light source, UK

0.1 sec exposure – for a bigger virus!

# A modern synchrotron beamline – 50 metres of optics



## Engineering driving science

A 'zoom lens' system to focus the X-rays –  
either at sample or on the detector

**$10^{12}$  X-rays per second**

**Beam size 5 – 50  $\mu\text{m}$**

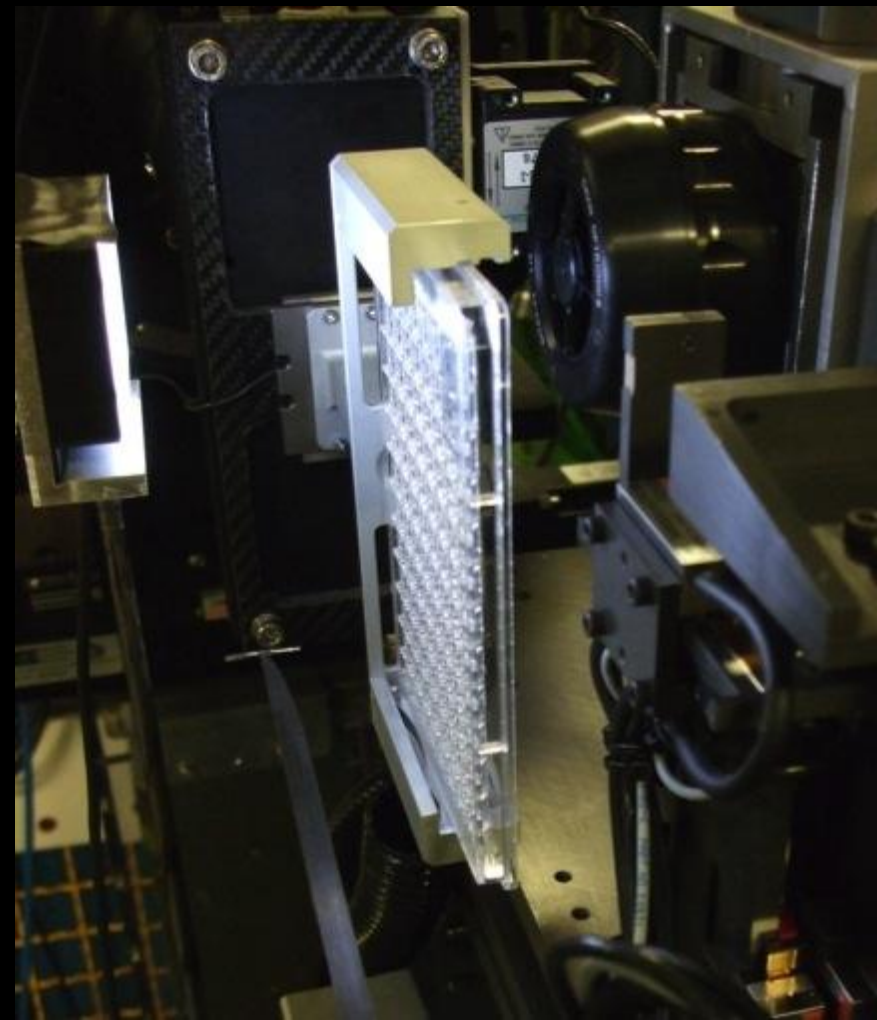
Gwyndaf Evans, Danny Axford,  
David Waterman, James Foadi,  
Jun Aishima, Robin Owen  
Diamond Light Source



So we have gone from taking long exposure photographs with a pinhole camera to high speed movies on a sophisticated camera with a zoom lens ... and there is more to come!

We can deal with fragile  
pathogenic crystals

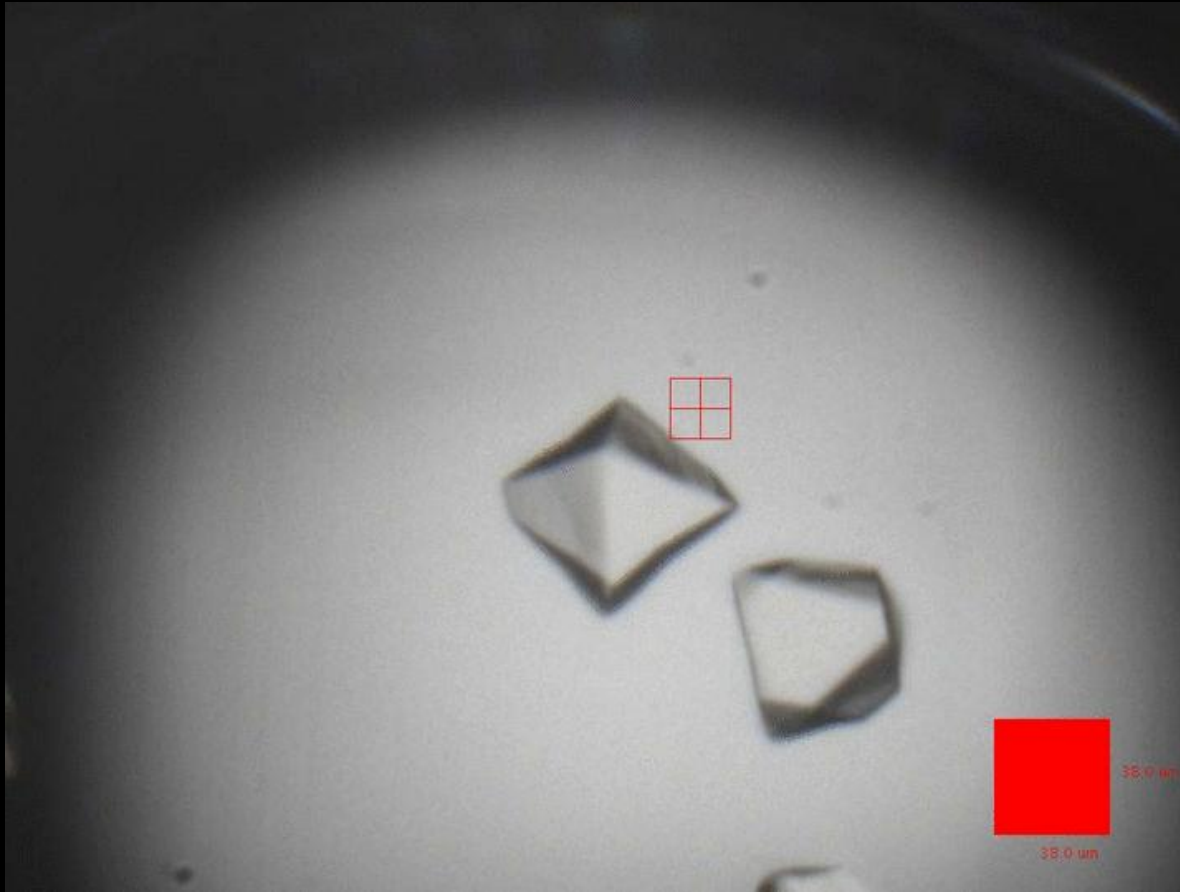
- *In situ* in the tiny (nl) drops in  
which they are grown



We can deal with fragile pathogenic crystals

Looking along the X-ray beam – each crystal survives 0.4s

– the X-ray beam is 1/50<sup>th</sup> of a millimetre across



124 staff, plus E Fry, JS Ren, A Kotcha DIS, Oxf (Axford et al 2012)

And there are revolutions in electron microscopy...

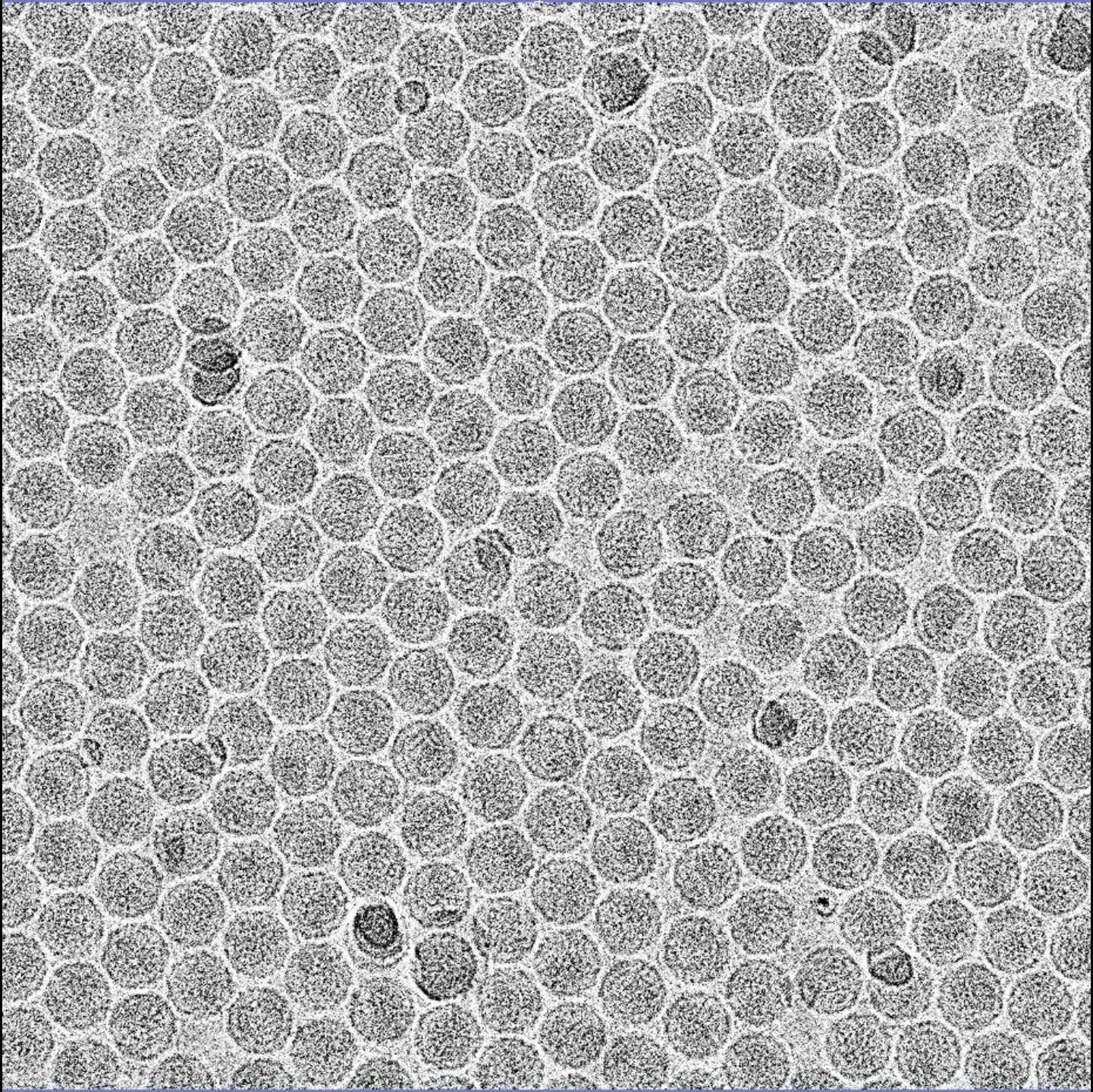
Wonderful new detectors, that collect 4k movies, and allow virus particle movements to be tracked in software ... so we can see atomic detail

Live virus in cryo – in a water 'glass'

Expensive – hence a National Facility for the UK

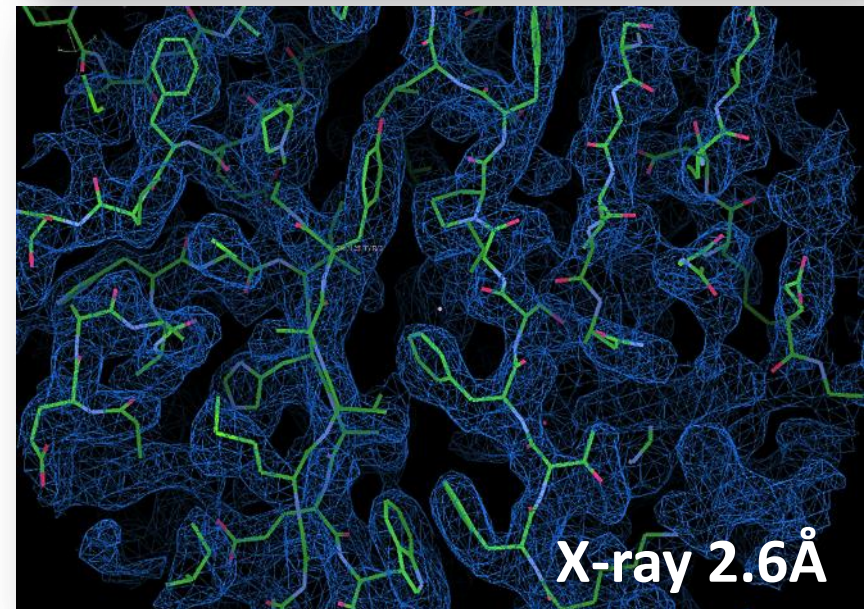
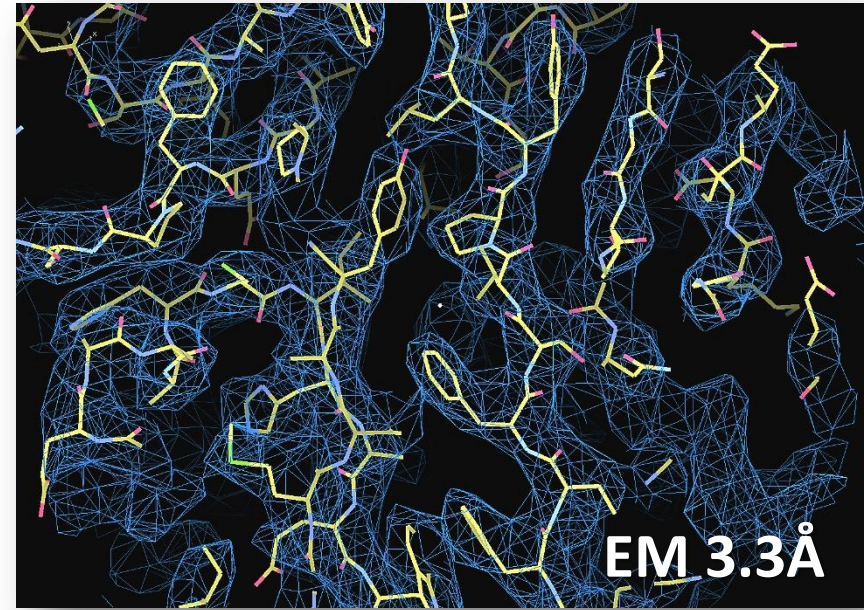
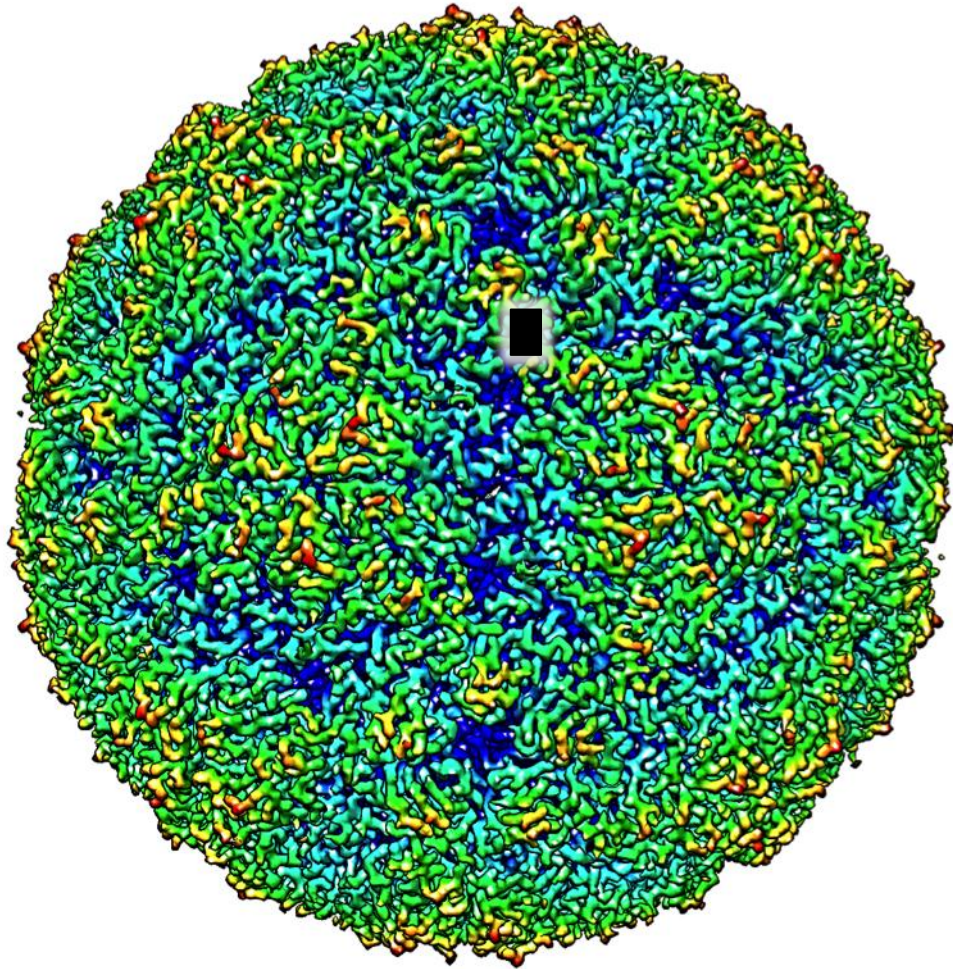








# Example from foot-and-mouth disease virus electron microscopy and X-ray structures



Both of these methods are now pretty quick (<1/2 day data collection), and require smaller amounts of sample than in the past

# Our targets: Picornaviruses

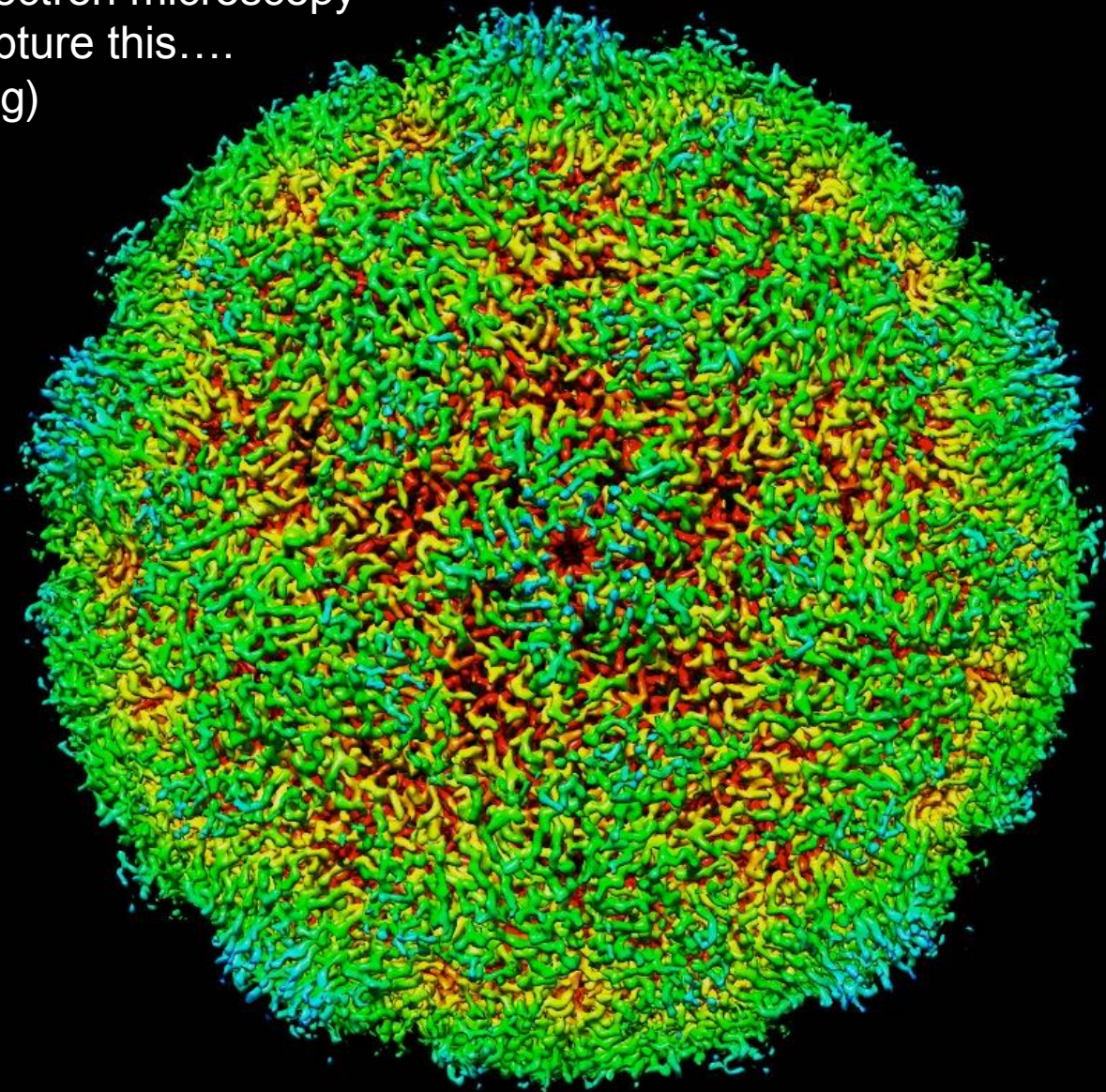
- RNA viruses with an icosahedral protein capsid – no lipid!
- name is derived from *pico*, meaning small, and RNA – *ie* small RNA virus
- include many important pathogens of humans and animals ranging from acute "common-cold"-like illnesses, to poliomyelitis, to chronic infections in livestock
- **We have a smattering of vaccines, based on 50 year old technology and NO licensed drugs.**
- **B cell responses critical – complete virus particle required for proper antibody protection**

# **Our targets: Picornaviruses**

- **Aim to do two ‘simple’ things**
  - **make a synthetic version**
  - **make it stable enough to do the job**
- **I will give as a major example Foot-and-mouth disease virus (FMDV) – but even in this family there are differences, for instance swine vesicular disease virus is more like poliovirus**

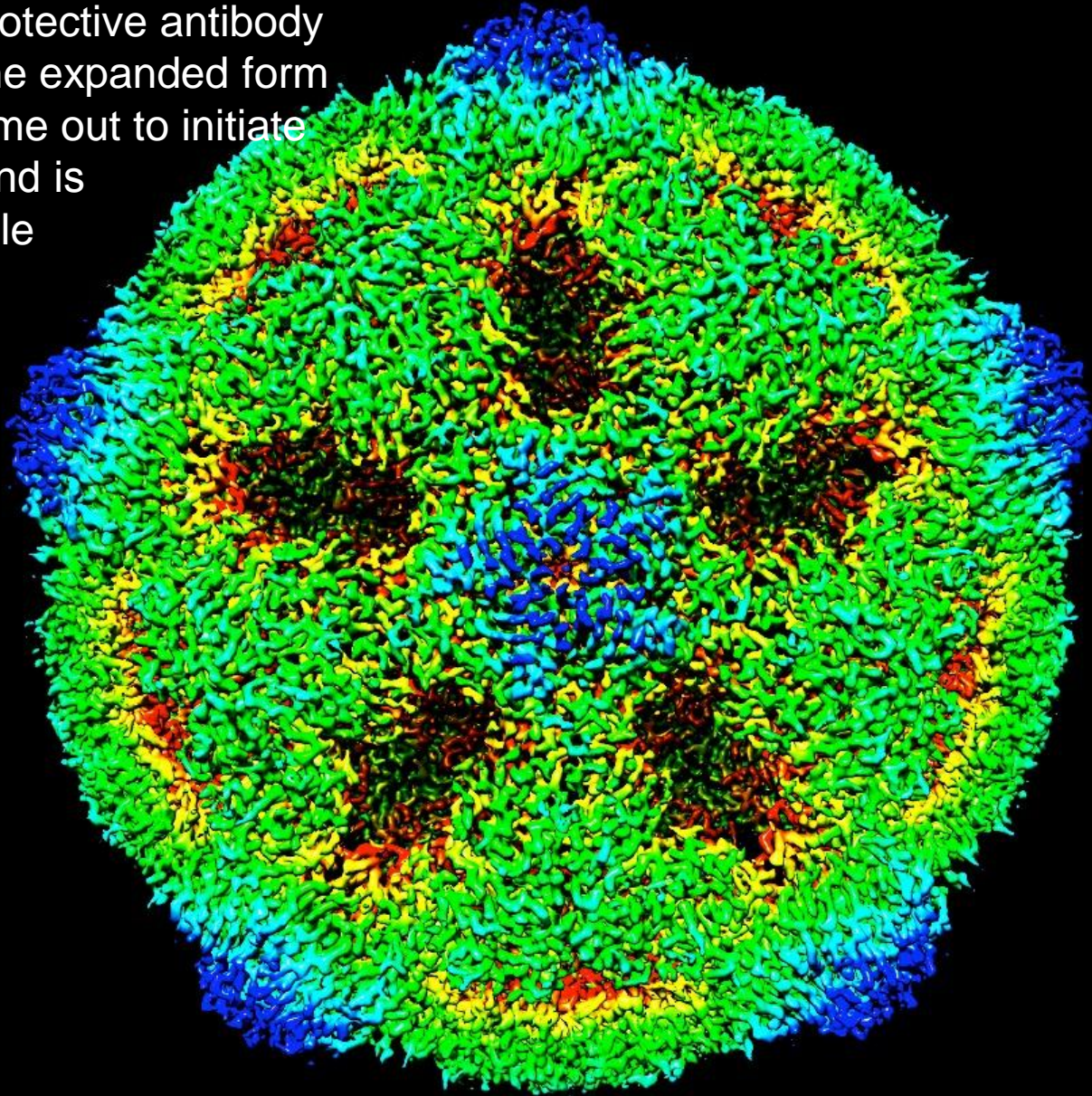


These enteroviruses 'breathe'  
A series of electron microscopy  
structures capture this....  
(Xiangxi Wang)





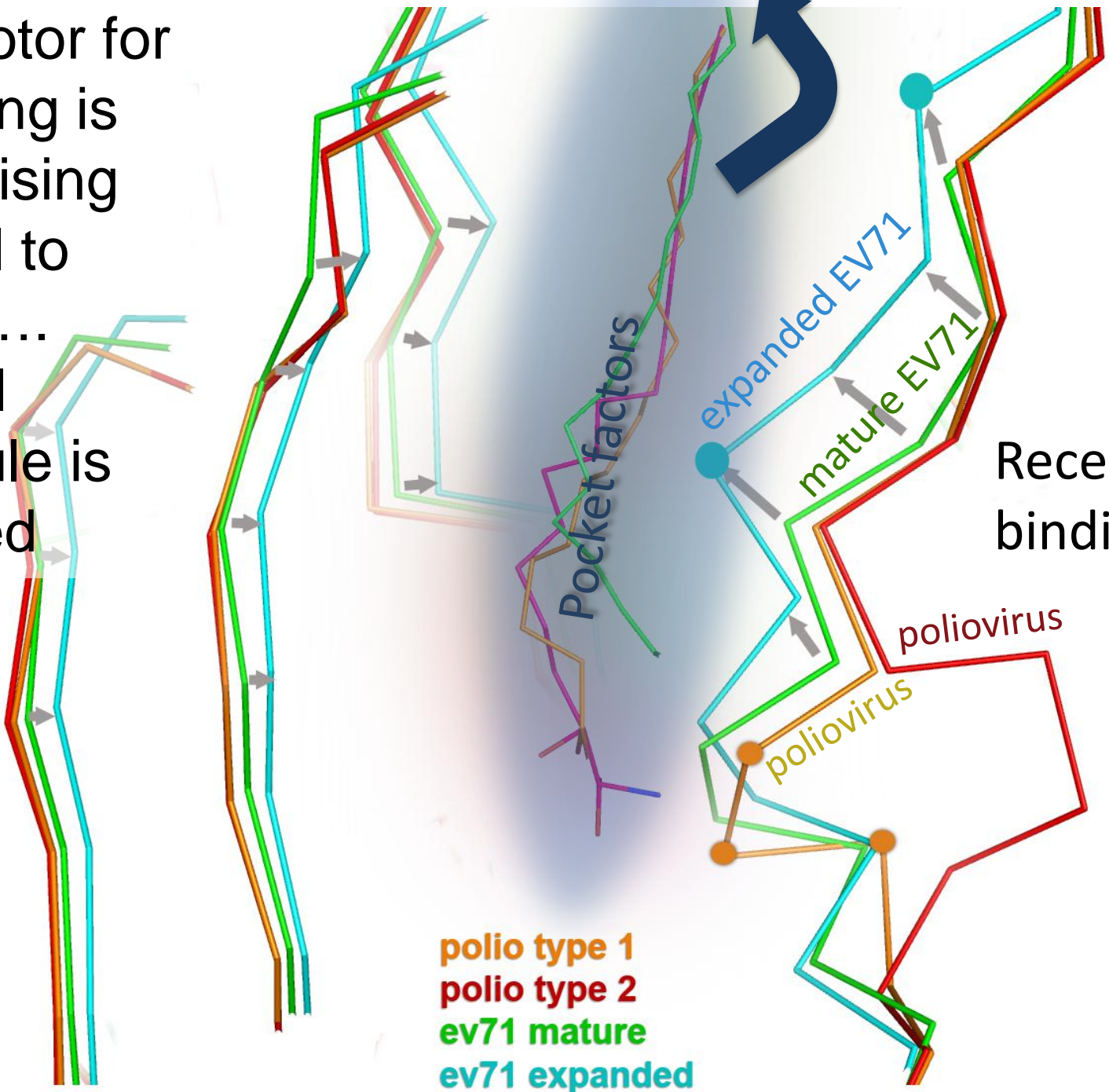
Only the unexpanded form can  
generate a protective antibody  
response – the expanded form  
lets the genome out to initiate  
infection ... and is  
the most stable  
**PROBLEM**  
since minus  
RNA stability  
is reduced



The motor for switching is recognising the cell to attack....

A small molecule is expelled

Inside



polio type 1  
polio type 2  
ev71 mature  
ev71 expanded

Receptor binding site

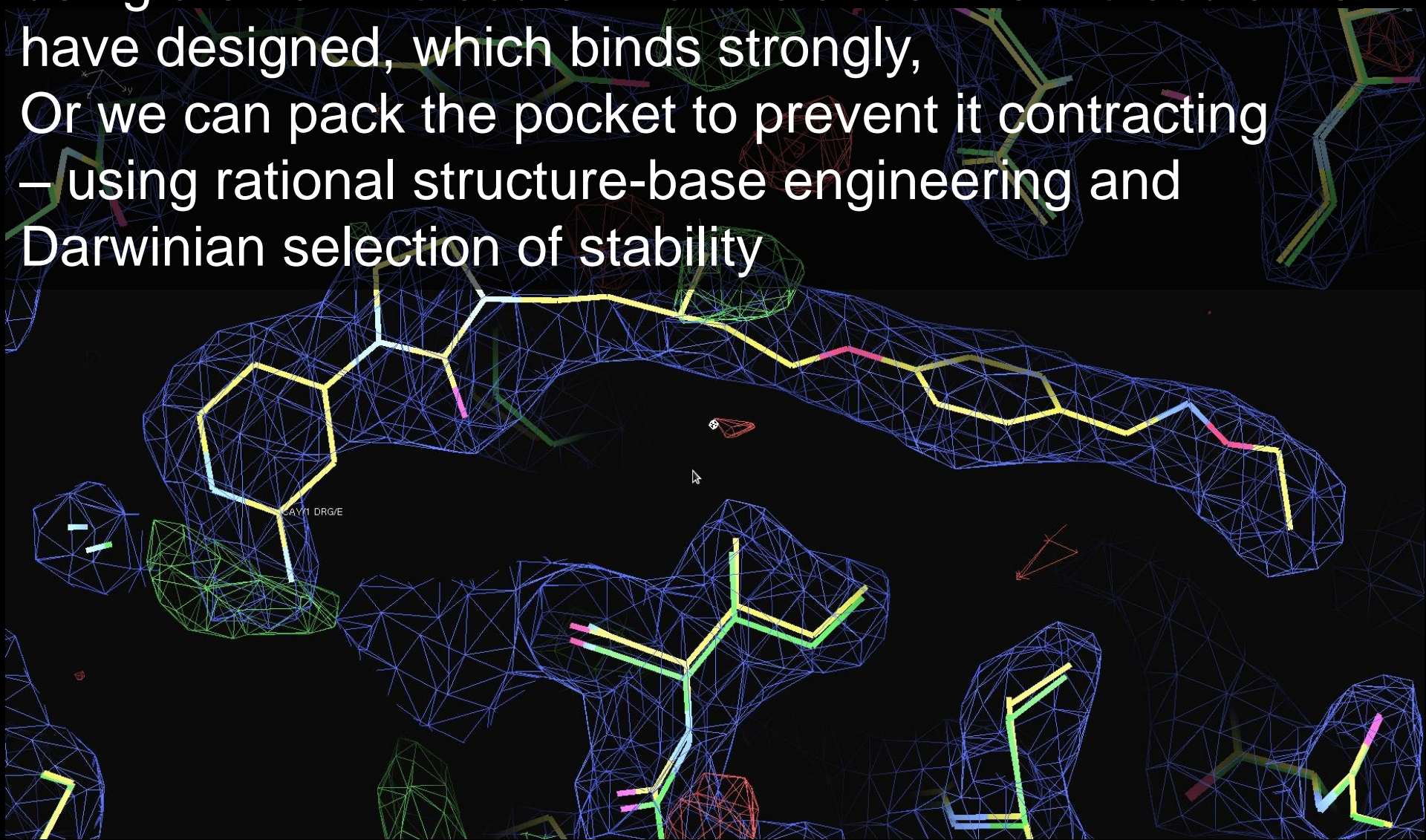
poliovirus  
poliovirus

expanded EV71  
mature EV71

Pocket factors



So we might lock the virus in the correct antigenic state using a small molecule – for instance this molecule we have designed, which binds strongly,  
Or we can pack the pocket to prevent it contracting – using rational structure-based engineering and Darwinian selection of stability



# Polio - collaborators

Collaborator in USA:  
James Hogle,  
Harvard Medical School



Polio stability  
Virus isolates  
Virology



Plant expression

Structure based  
prediction  
Pocket binding  
drugs



Polio stability  
N/H antigen  
status

Baculovirus  
expression  
VLP synthesis



University of  
**Reading**

WHO – Gates funding



- **For FMDV we can use a different approach ... as we will see ...**



# Foot-and-mouth disease – structure-based vaccine development

Supported by

**wellcome**trust



Liz Fy, Abhay Kotecha, Ren Jingshan, Claudine Porta (Pirbright), Tom Walter, Karl Harlos



Robert Esnouf

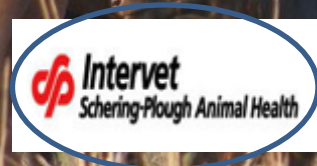


Bryan Charleston, Julian Seago, Terry Jackson, Alison Burman, Clare Grant, John Hammond



Ian Jones, Reading University

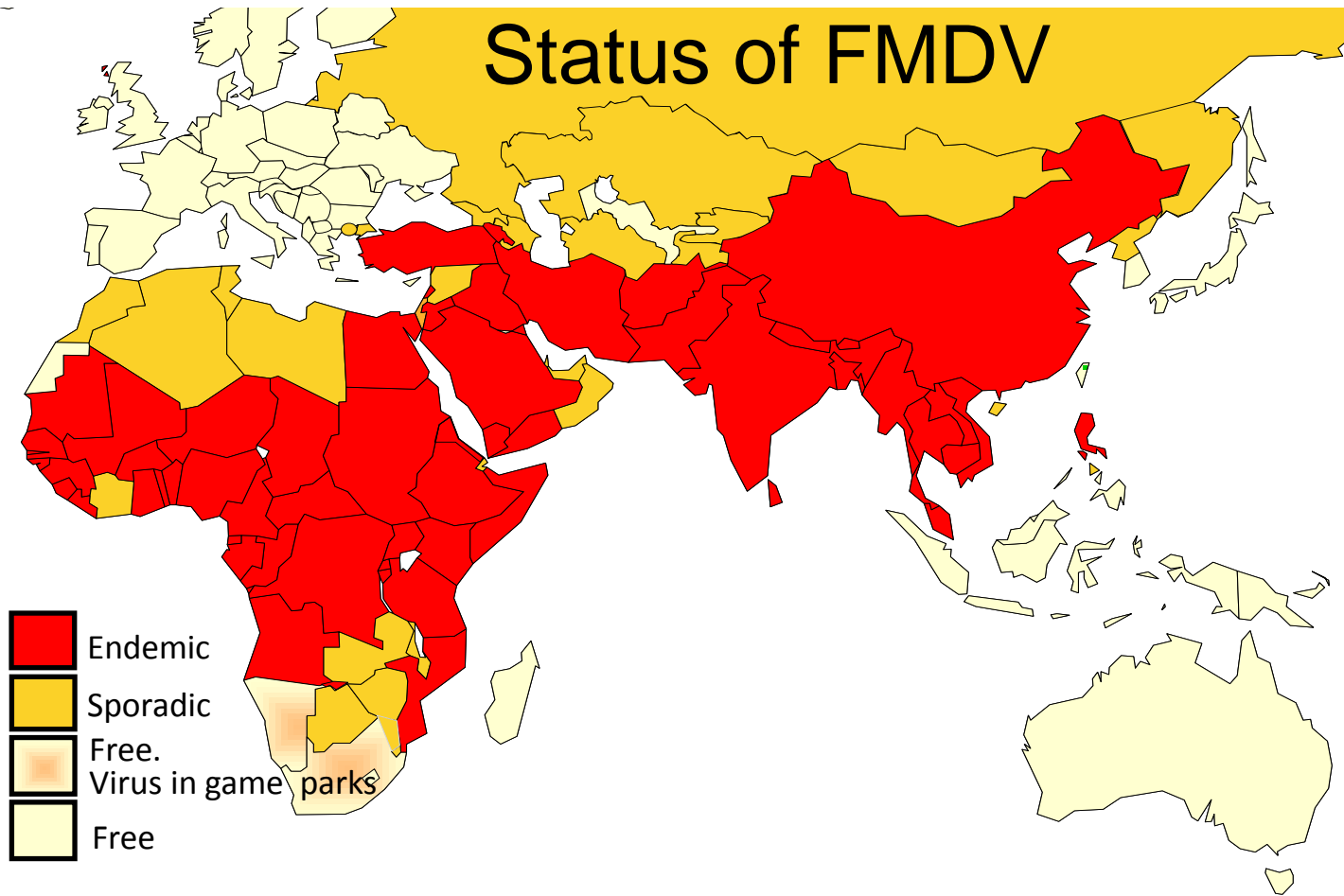
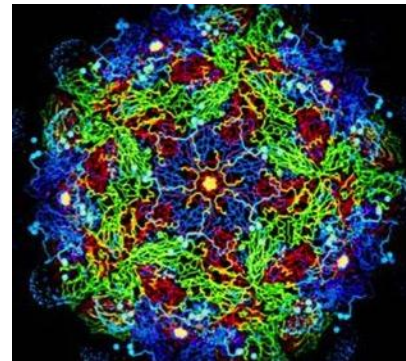
Francois Maree, Katherine Scott, SA Dr Venkat, Bangalore



MSD Intervet

# Foot-and-mouth disease.

A global food security issue  
and impediment to wellbeing





- Much of the global FMD burden of production losses falls on the world's poorest communities, and those which are most dependent upon the health of their livestock.
- Overall the direct losses **limit livestock productivity** creating a **food security** issue and contributing to **malnutrition**



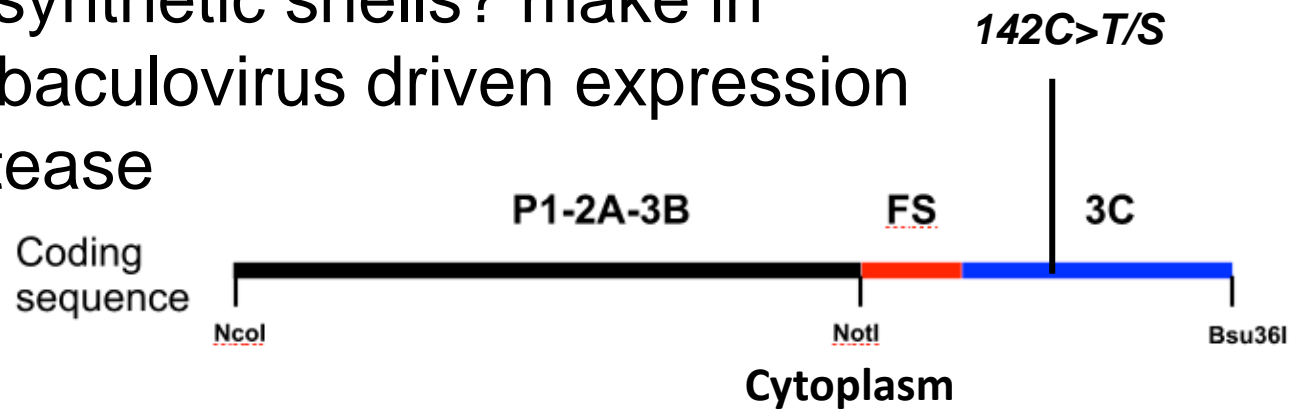


- Much of the global FMD burden of production losses falls on the world's poorest communities, and those which are most dependent upon the health of their livestock.
- Overall the direct losses **limit livestock productivity** creating a **food security** issue and contributing to **malnutrition**
- **Current vaccines** are inactivated live virus formulated with adjuvant
- Globally there is a **shortage of vaccine** – billions of doses needed.
- **This is 50 year-old technology**
- We are trying to bring this up-to-date: safe, cheap, effective (for some types of FMDV there is NO effective vaccine)
- (i) **How to make a synthetic virus-free vaccine?**
- (ii) **How to make it stable enough to be effective?**



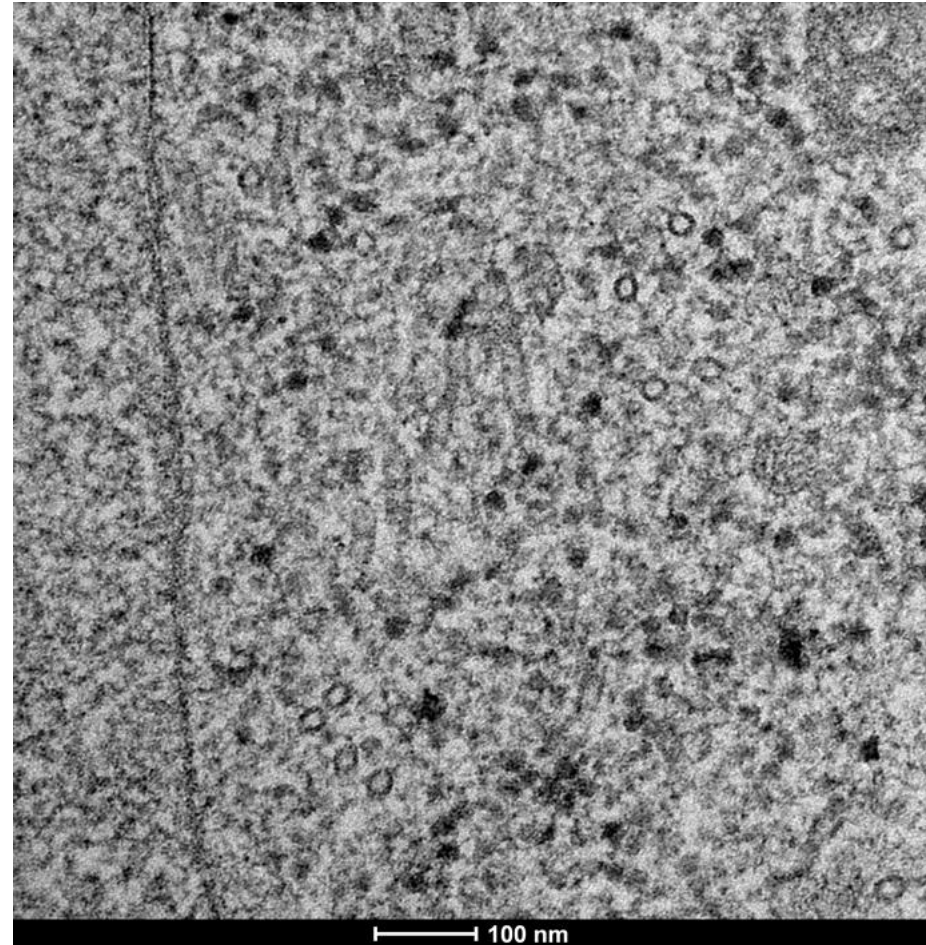
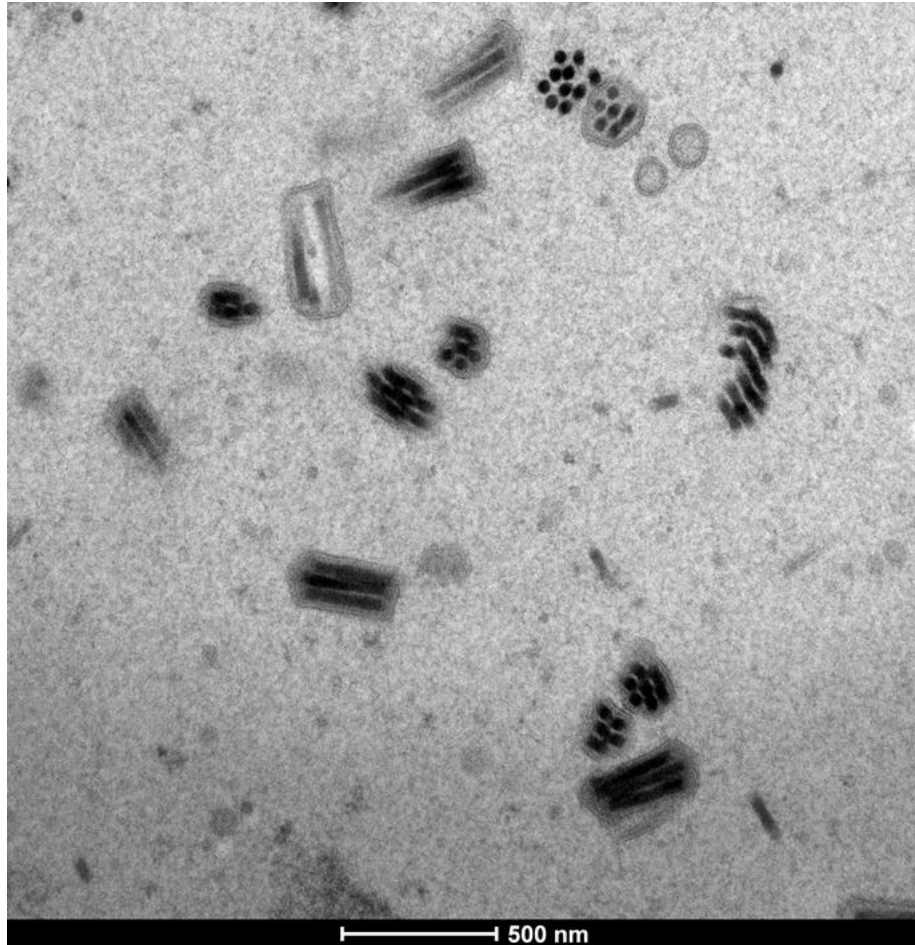
Q1: can we make synthetic shells? make in insect cells, using baculovirus driven expression – detune toxic protease

(Ian Jones, Reading)



Nucleus

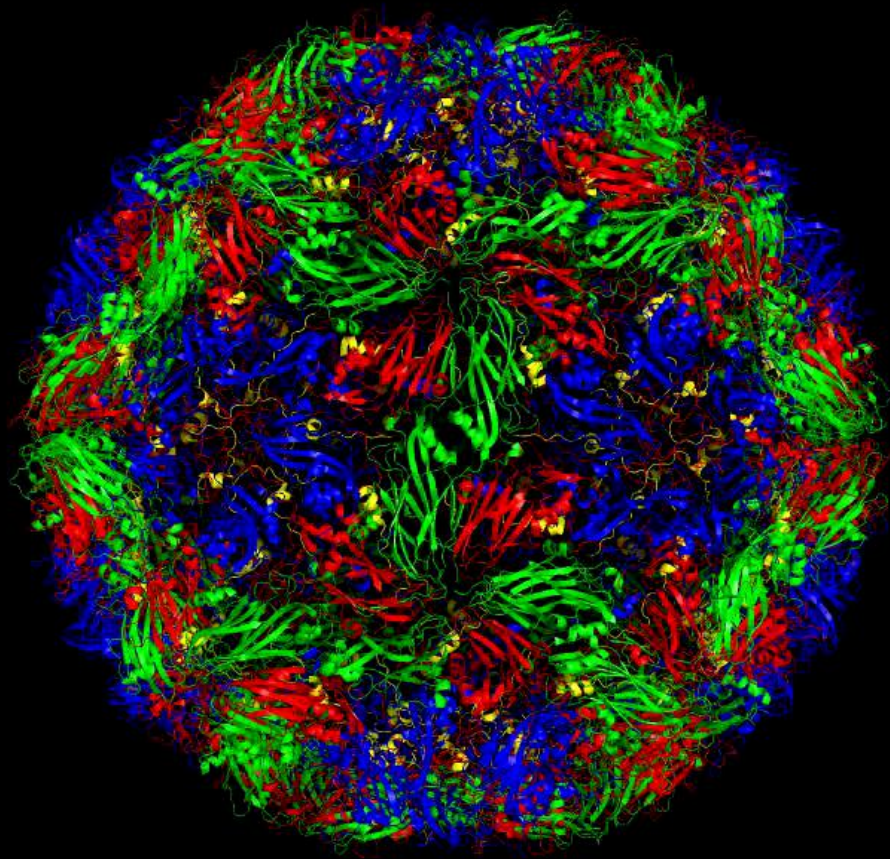
Cytoplasm





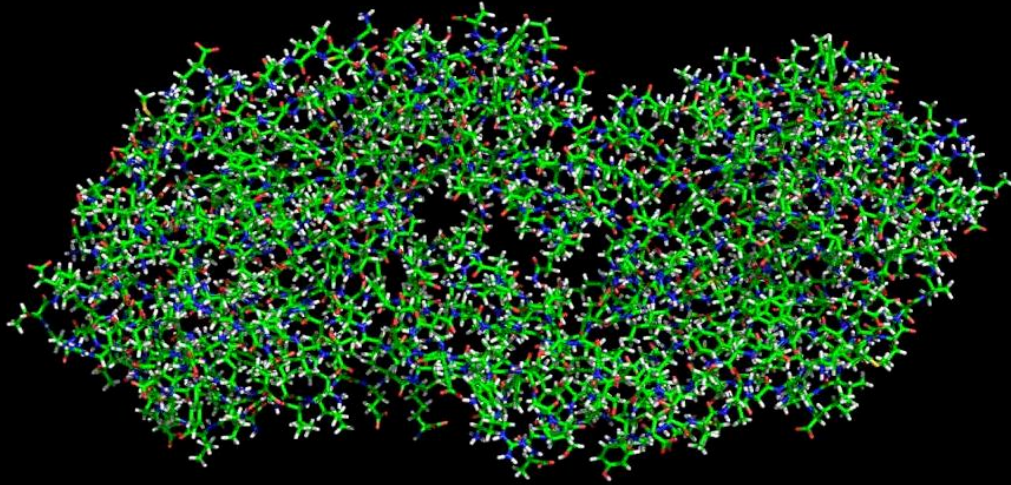
- Q2: Can structure help us re-engineer the particles to make them more stable?

FMD vaccines are fragile - capsids are unstable



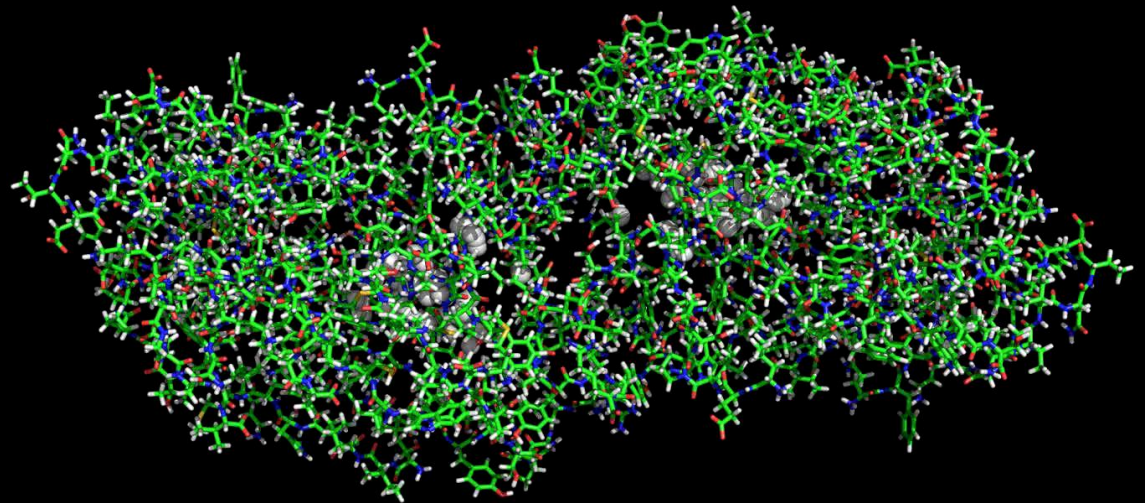
- Use *in silico* modelling to help solve the problem of what to re-engineer
- We first cut out the part that is the weak link ...





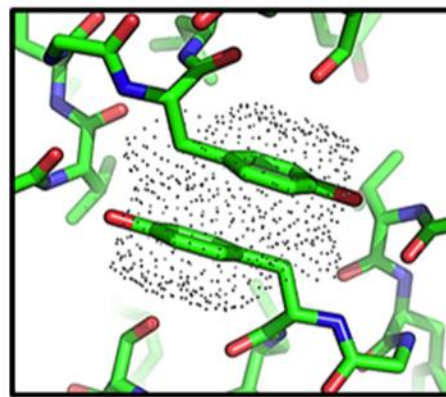
“Textbook”  
simulation –  
noise dominates

Modified  
simulation  
- target the  
weakspot

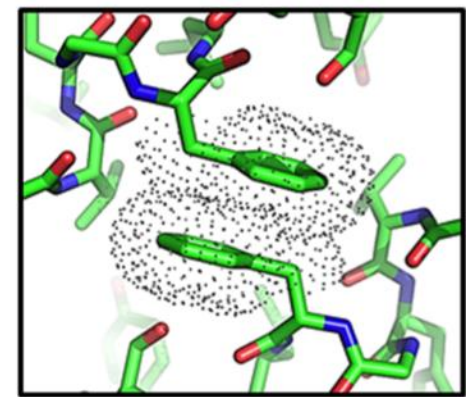


- We designed a series of single point mutations to stabilise the symmetry point at the centre of the interface between pentamers

and the predicted  
structures....

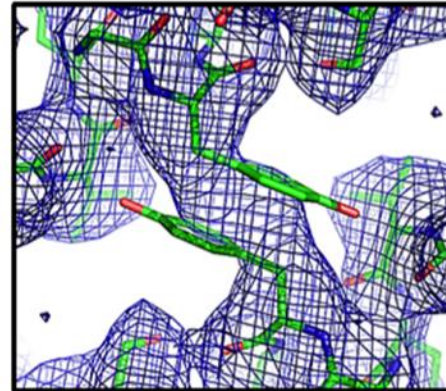


O1M VP2 S93Y

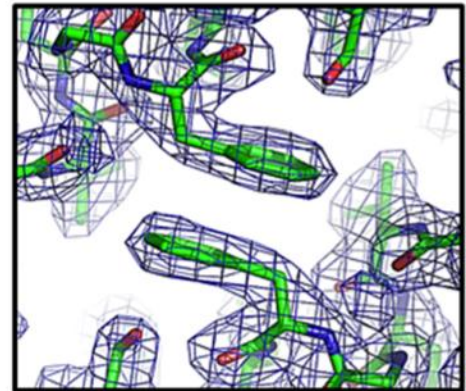


O1M VP2 S93F

look like the X-ray...



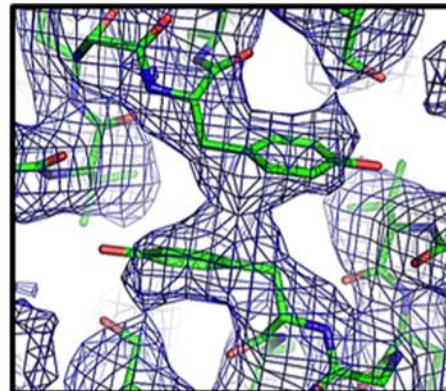
O1M VP2 S93Y



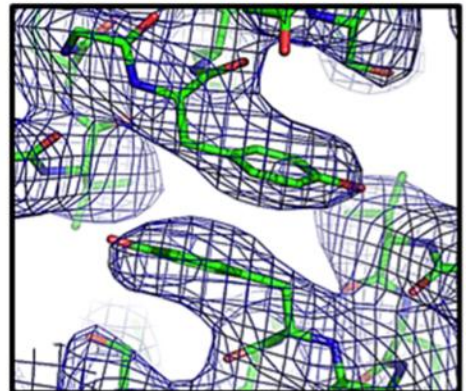
A22 VP2 S93F

and EM structures....

for two of the most  
unstable serotypes,  
O and SAT2



O1M VP2 S93Y

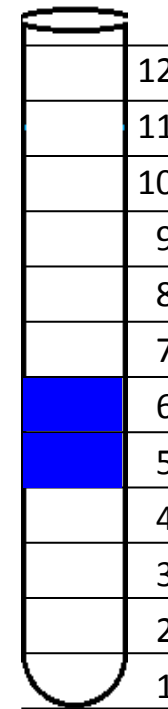
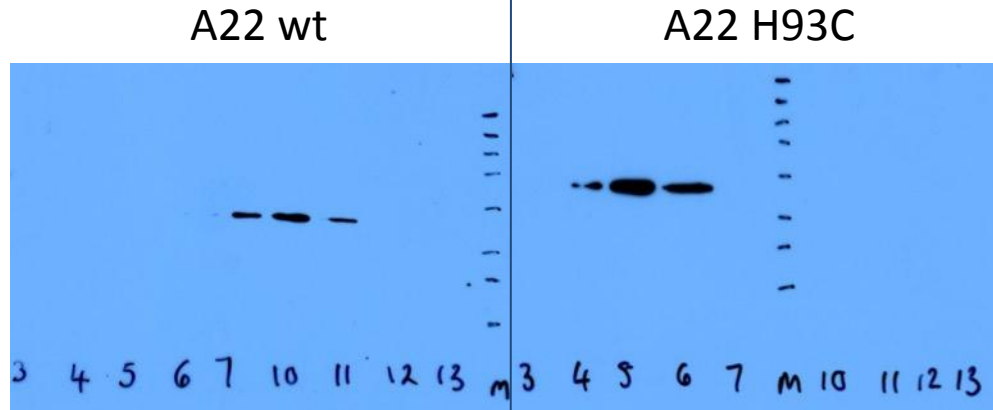
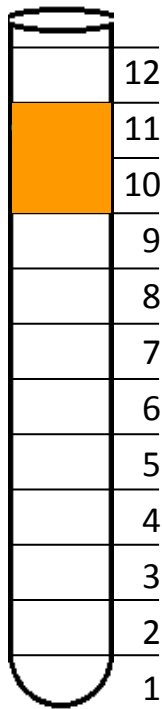
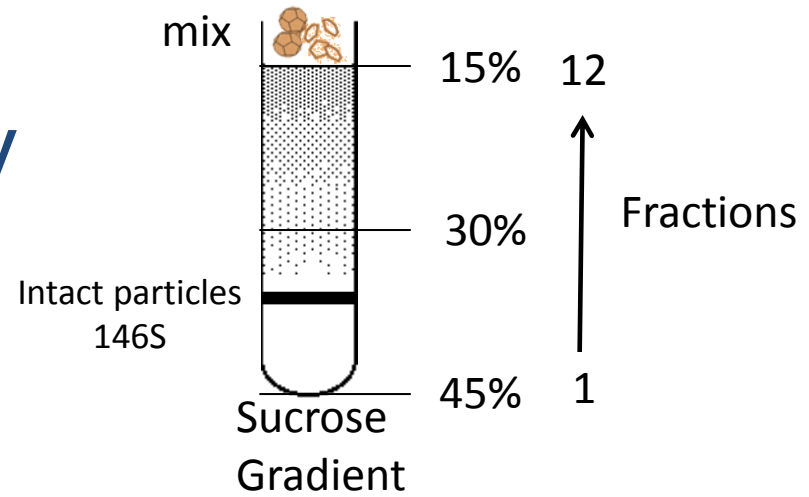


SAT2 VP2 S93Y



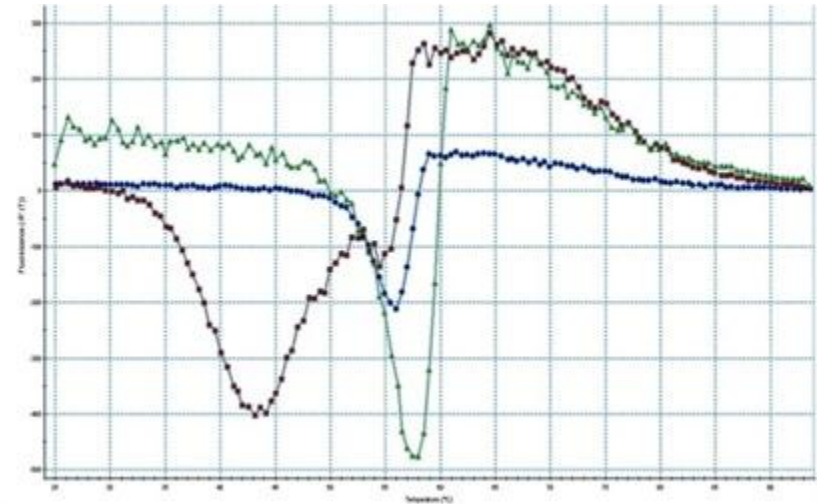
# Validation: Temperature stability

Heating at 56°C for 2 hrs



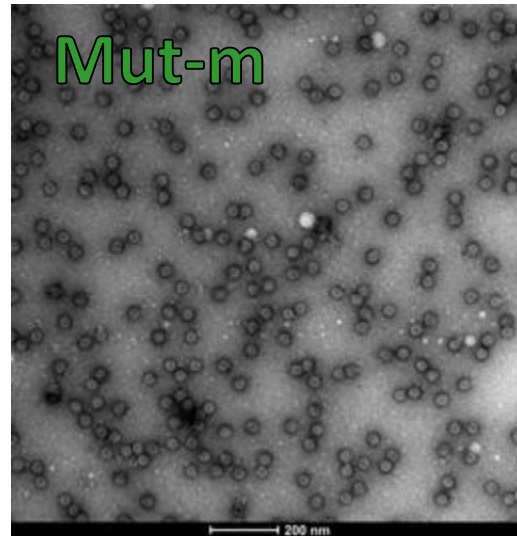
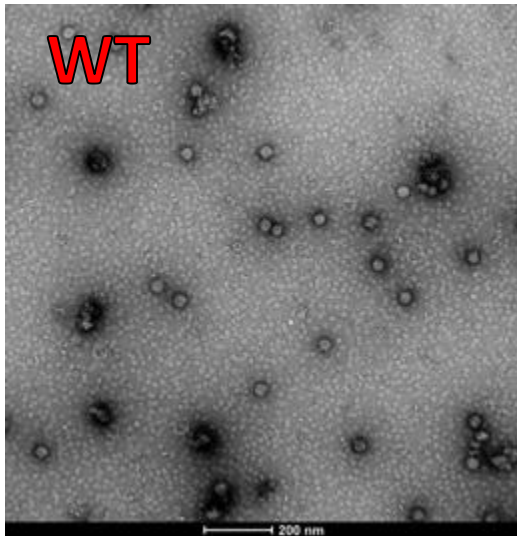
and they show *in vitro* stability (O serotype example)

Mutant	$\Delta\Delta G_{PB}$	Results
i	-5kcal/mol	Capsids stable
l	-3kcal/mol	Capsids stable
m	-5kcal/mol	Capsids stable
o	-3 kcal/mol	Few capsids, stable
p1	-2 kcal/mol	Not stable
p2	-3 kcal/mol	Poor yield & stability
p3	-.3 kcal/mol	Poor yield & stability



■ Wt (43°C)  
■ Mut-m (57°C)  
■ A22 (56°C)

**Thermoflour assay**

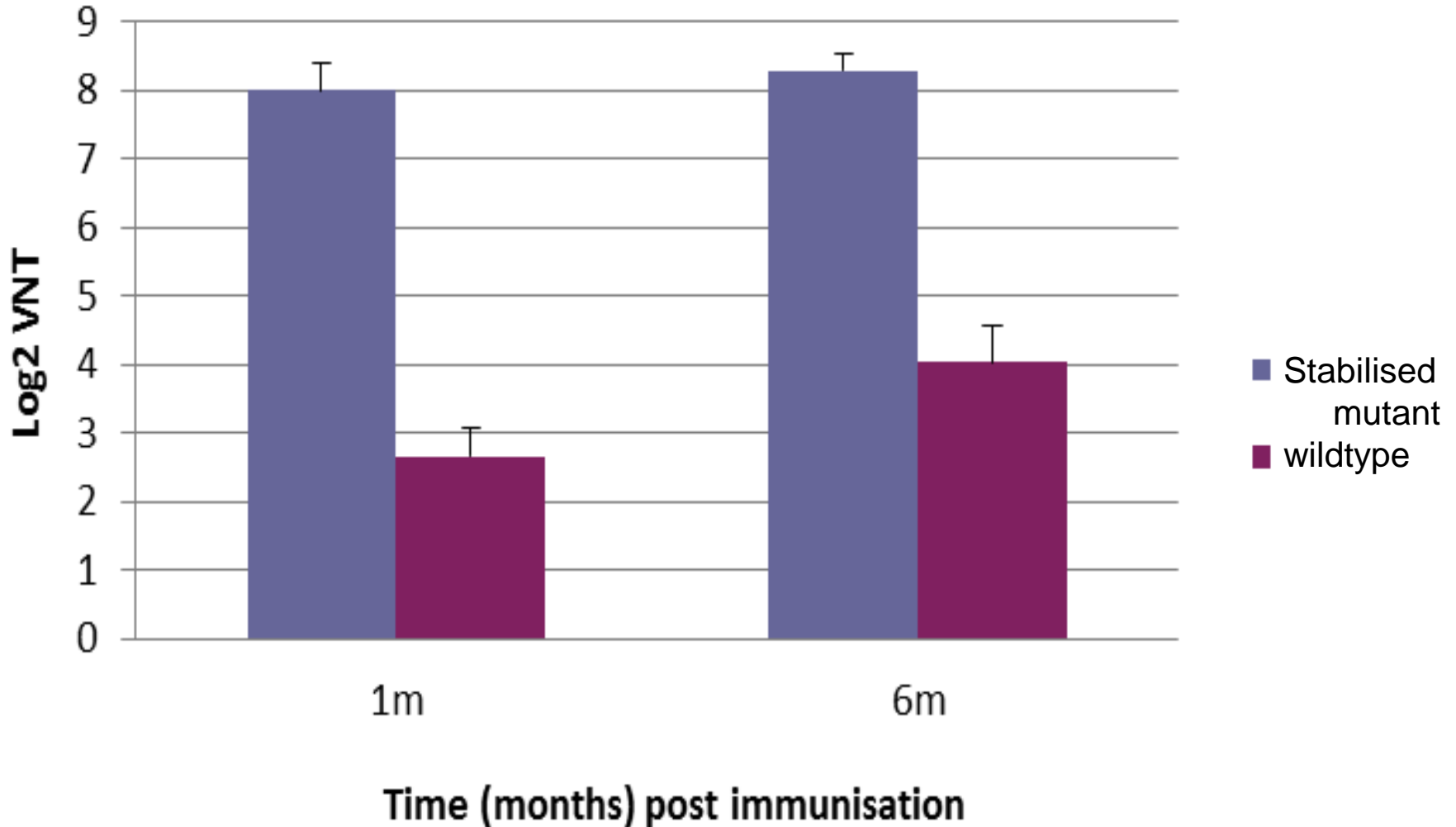


Purified capsids after 10 days in PBS at 4°C

Inactivated WT largely dissociates into pentamers

... and protect animals (SAT2 serotype example)

## FMDV SAT2 VNT titres





# FMDV Stabilised empty capsids, summary

- Improved storage characteristics
- Safe production - no live virus required
- Smaller production plants could be built locally altering the economics of vaccine production
- Vaccine can be quickly produced to new virus variants
- Simple diagnostic to discriminate vaccinated and infected animal
- Further animal tests and commercial viability tests underway

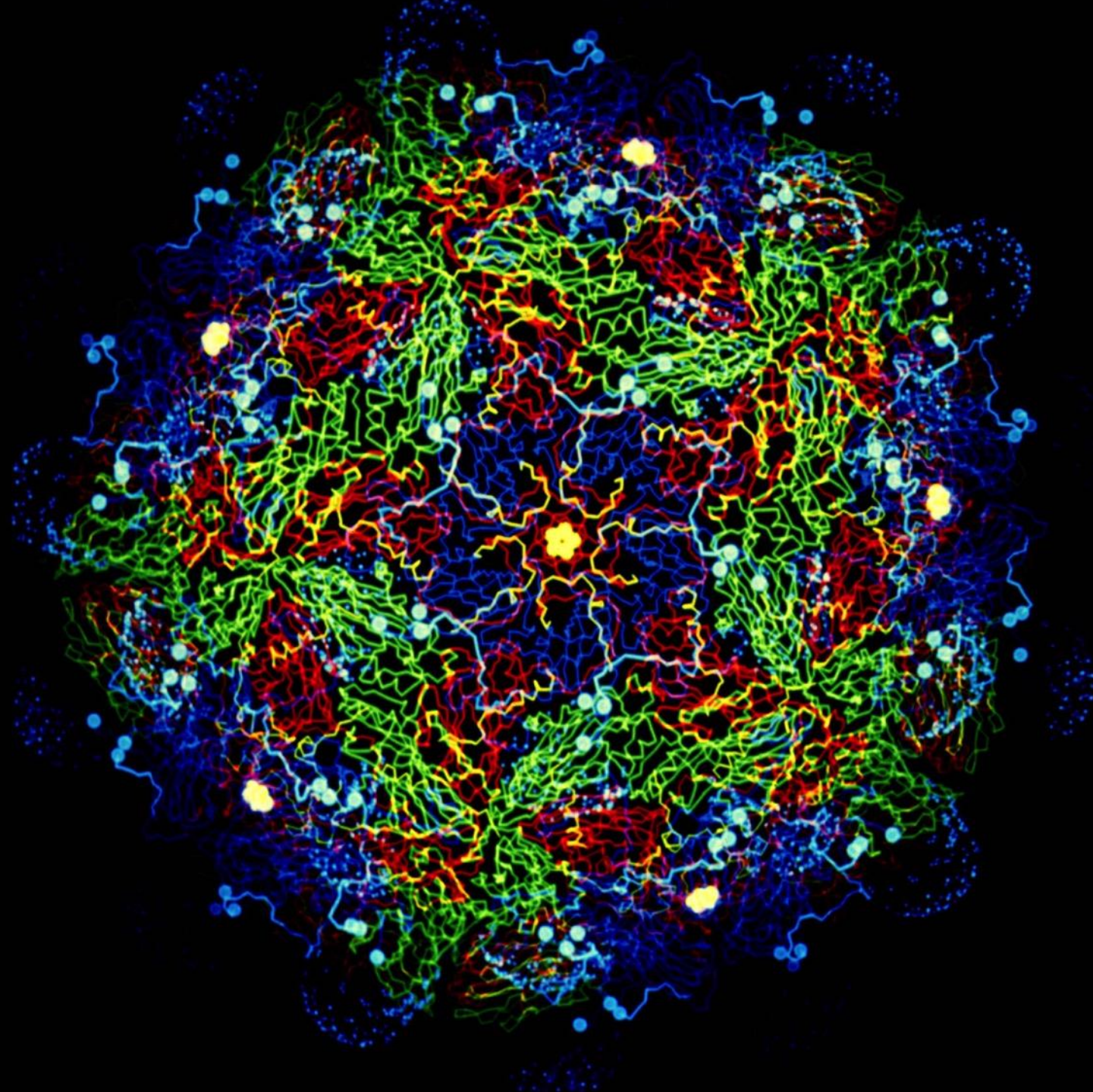
But can we think about making more cross-reactive responses?

Do we understand mechanisms for neutralisation?

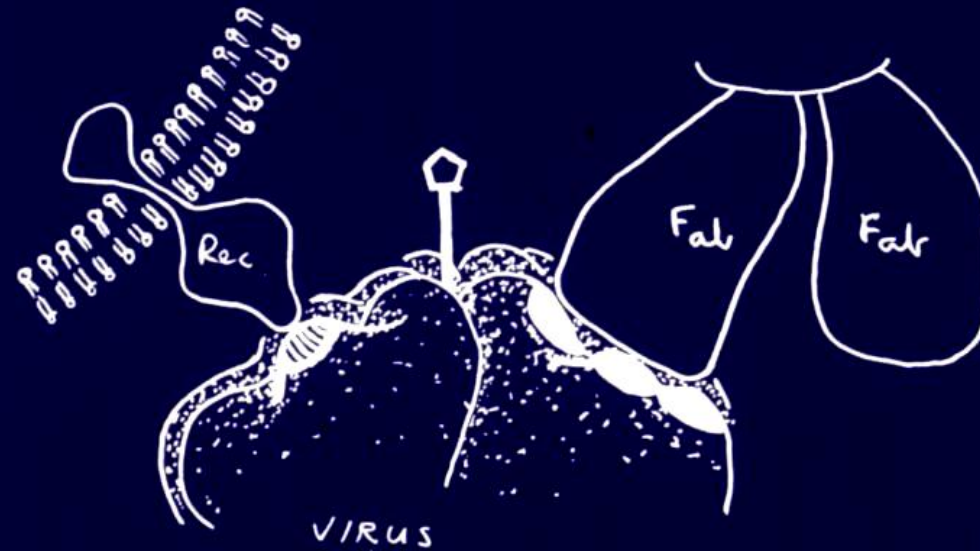
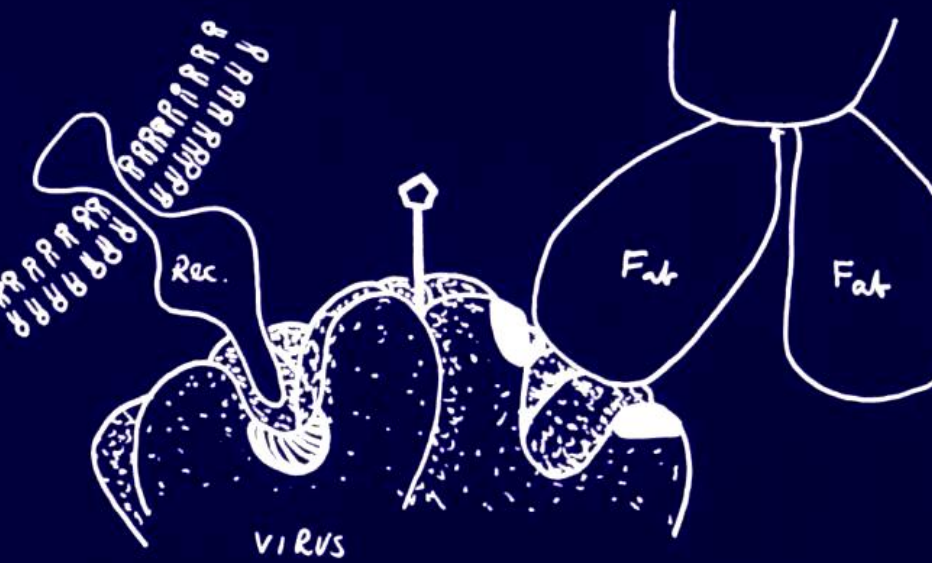
It appears that, although all of the virus surface should be visible there is one specific loop that seems to dominate.

The famous VP1 or 'FMDV loop'

The mechanism of neutralisation here is probably very straightforward – the loop binds the internalisation receptor – the  $\alpha\beta 6$  integrin.

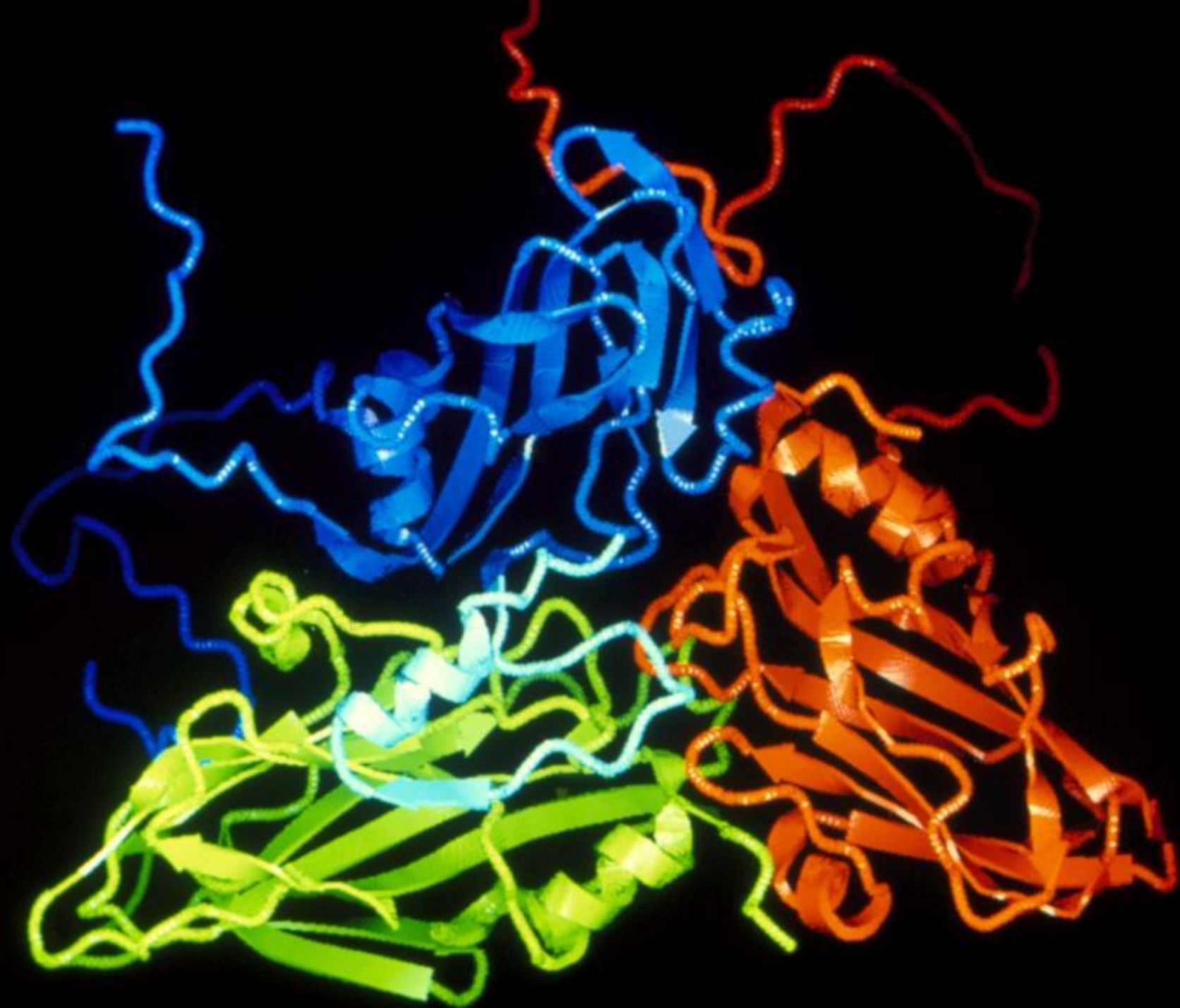


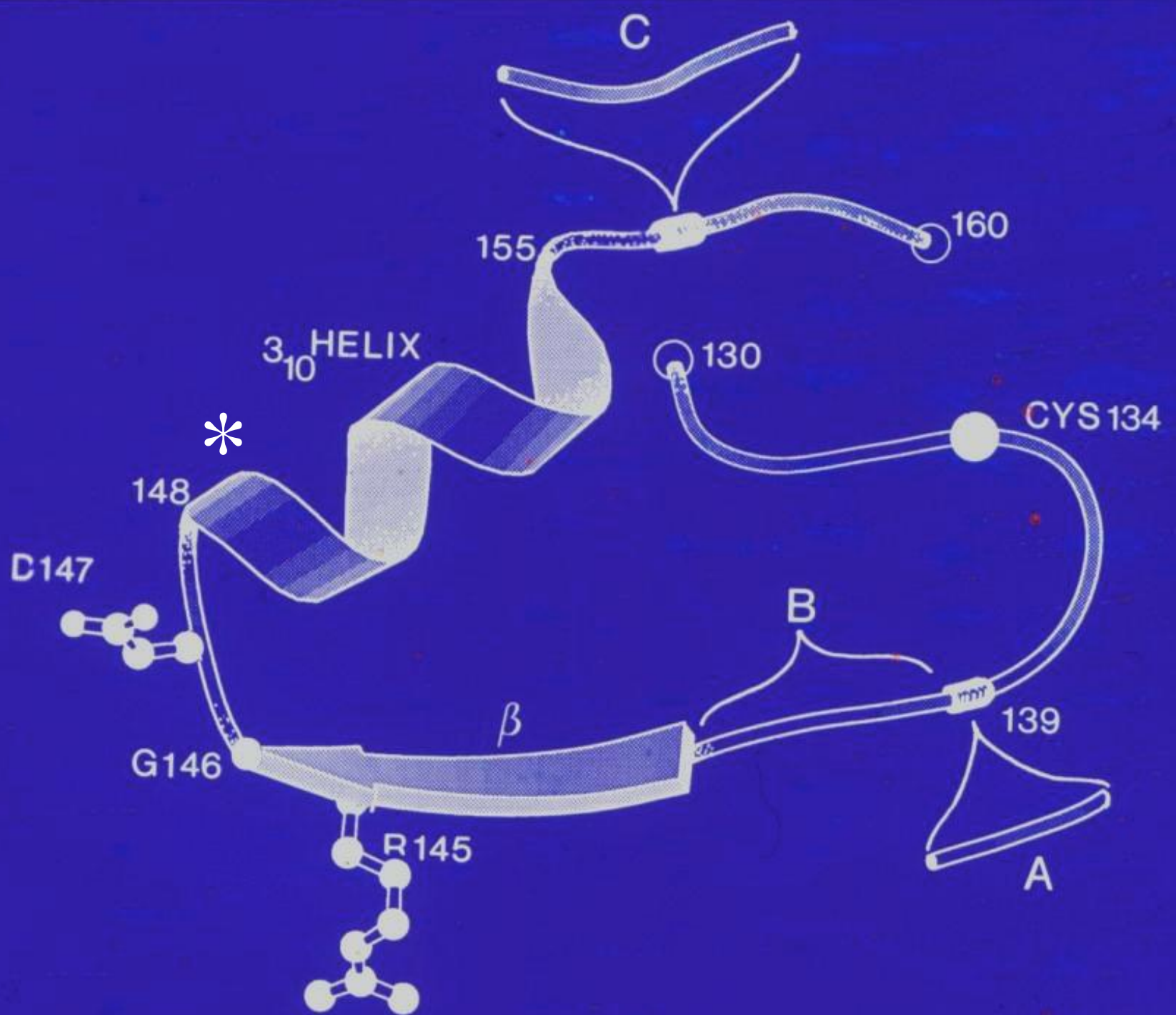




The Canyon Hypothesis

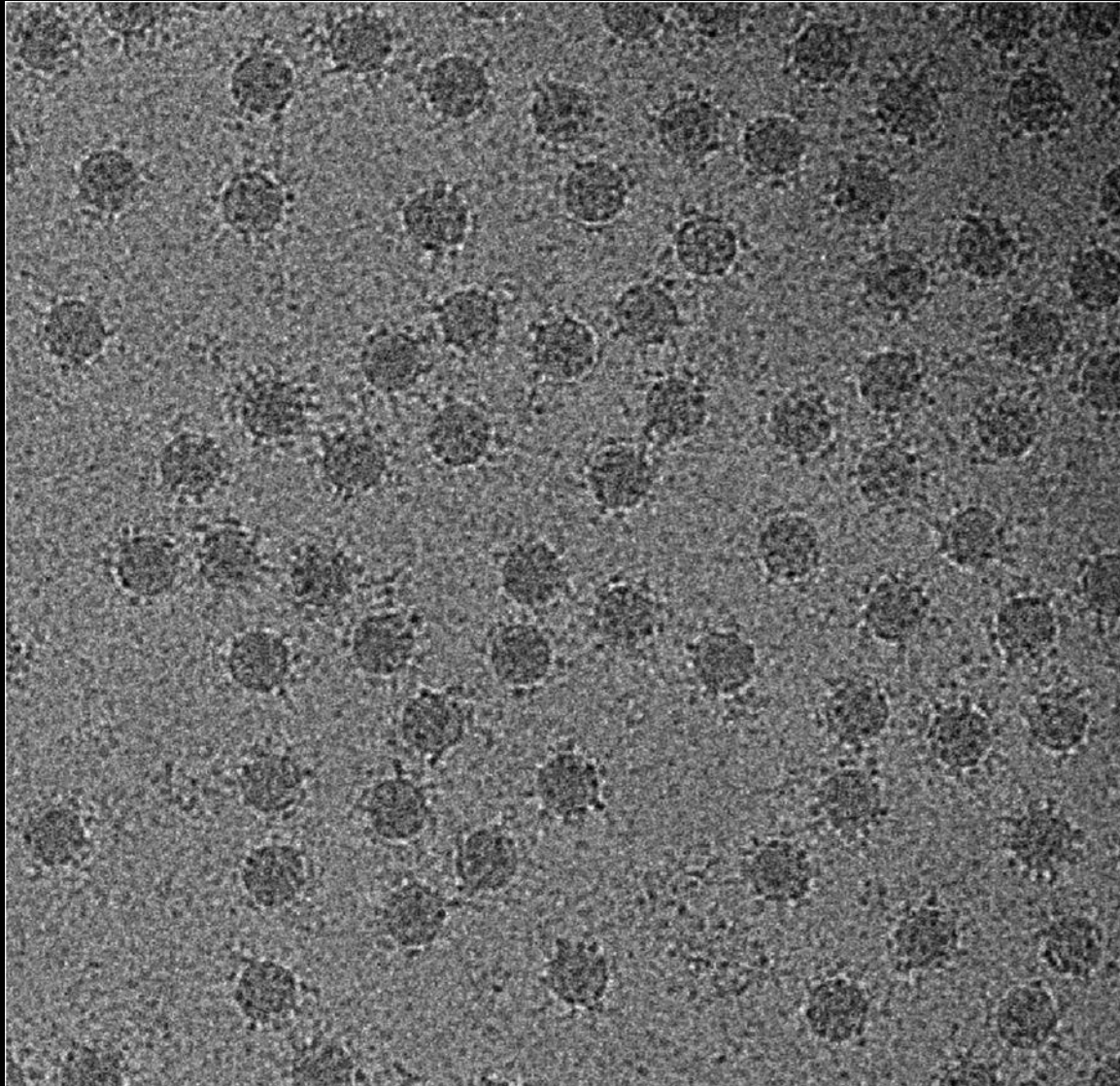
The FMDV Method







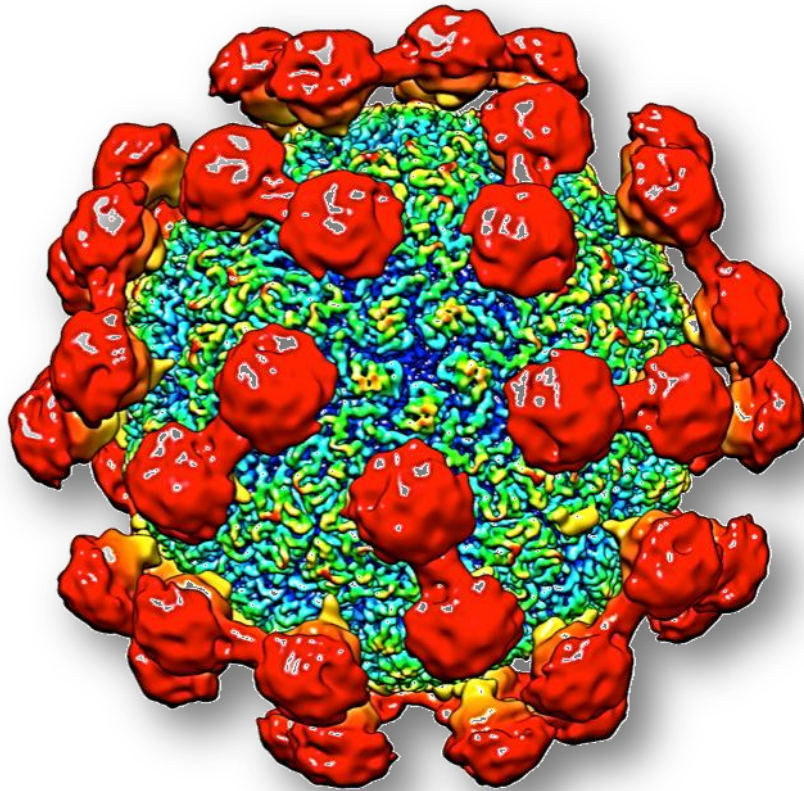
# FMDV – $\alpha\nu\beta 6$ complex + 2mM Mn



small headpiece  
No Hybrid domain

# O1M – $\alpha\nu\beta 6$ complex

(collaboration with Prof. Tim Springer, Harvard)

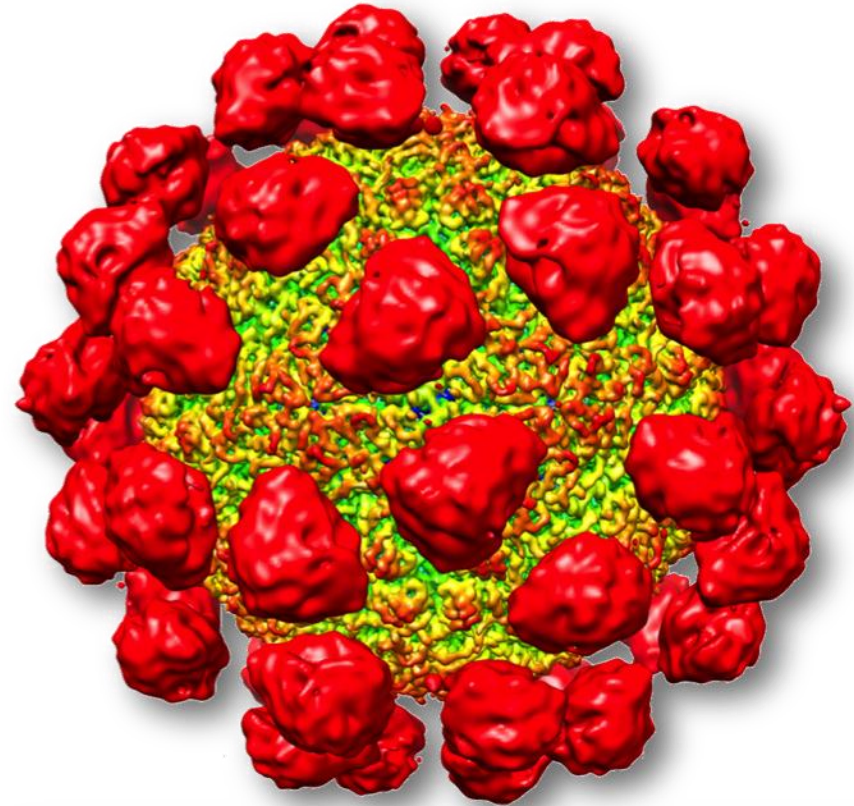


10900 Particles

Capsid resolution 3.4Å

Integrins are flexible - filtered at 10Å

2mM Mn

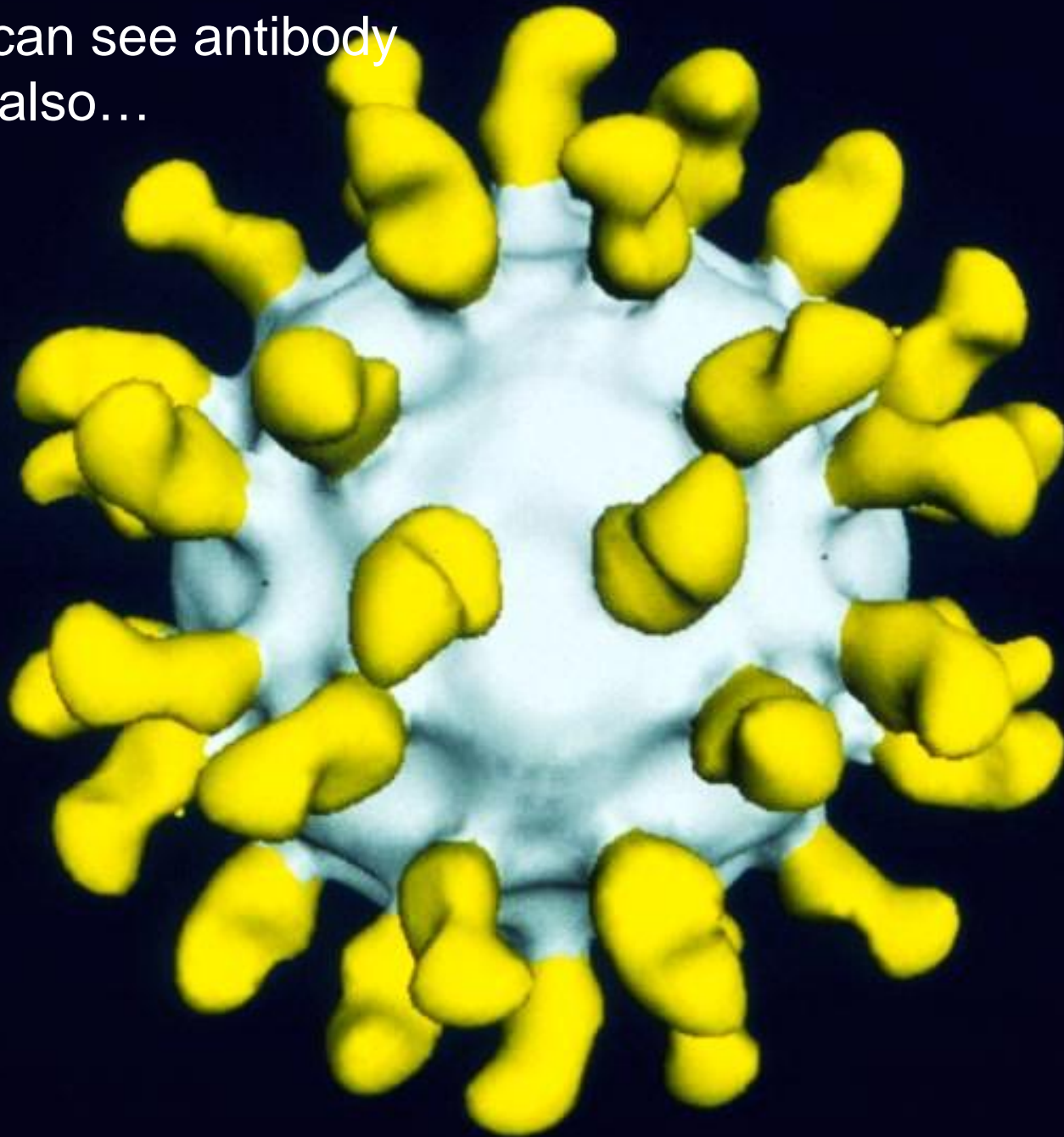


1649 Particles

Capsid resolution 3.4Å

Integrins are flexible - filtered at 10Å

But we can see antibody  
binding also...





We (Bryan Charleston leading) have set up a pipeline to investigate the immune response to

Deep IgG sequencing, to follow changes with vaccination (delta T, delta serotype)

Bioinformatics

Selection of Abs

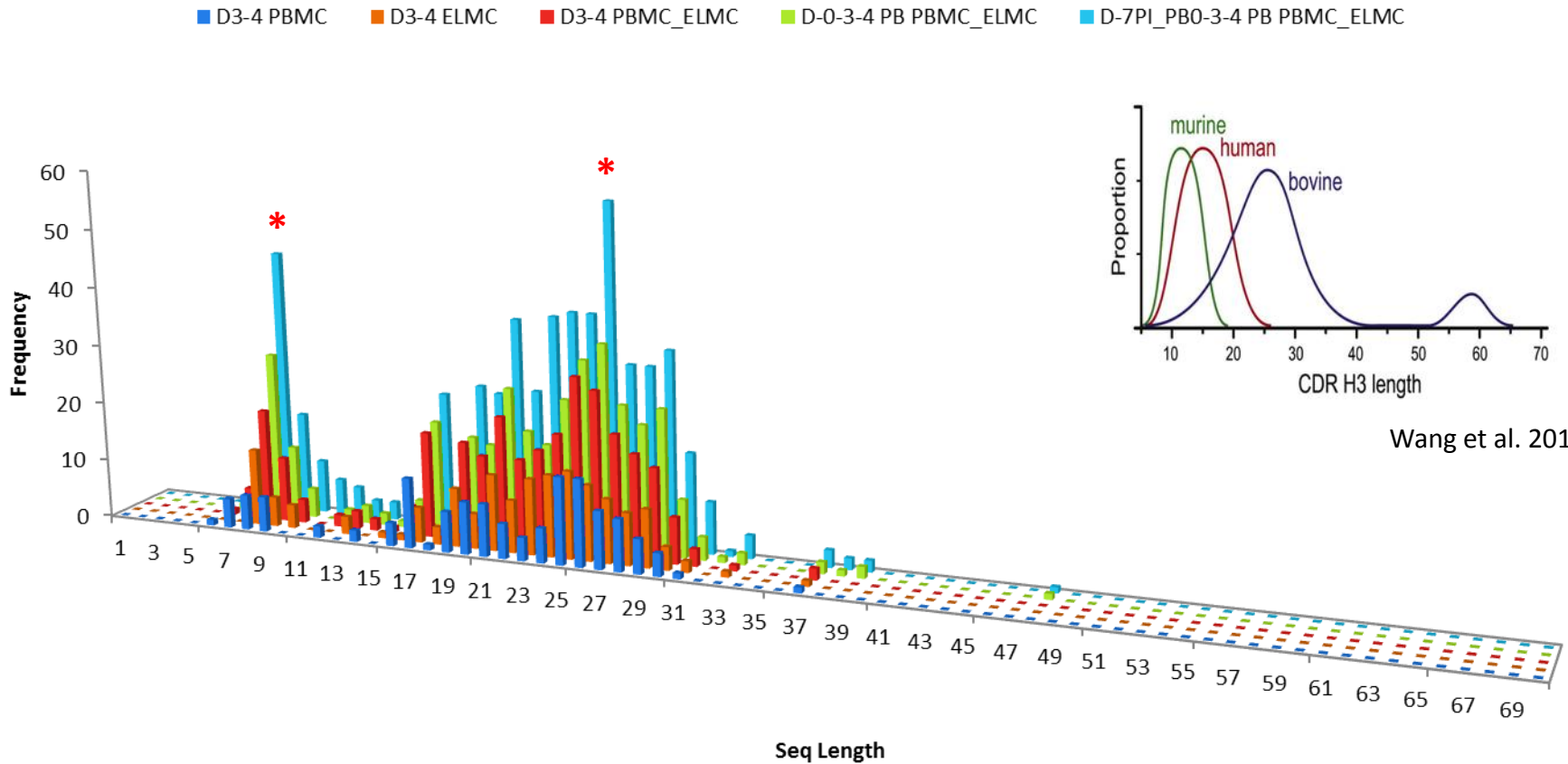
Synthetic gene blocks

Express / Purify

Ag Binding?

Structure Ab (X-ray) and Ab/Ag (EM)

# Distribution of CHD3 length of Vh Post Boost

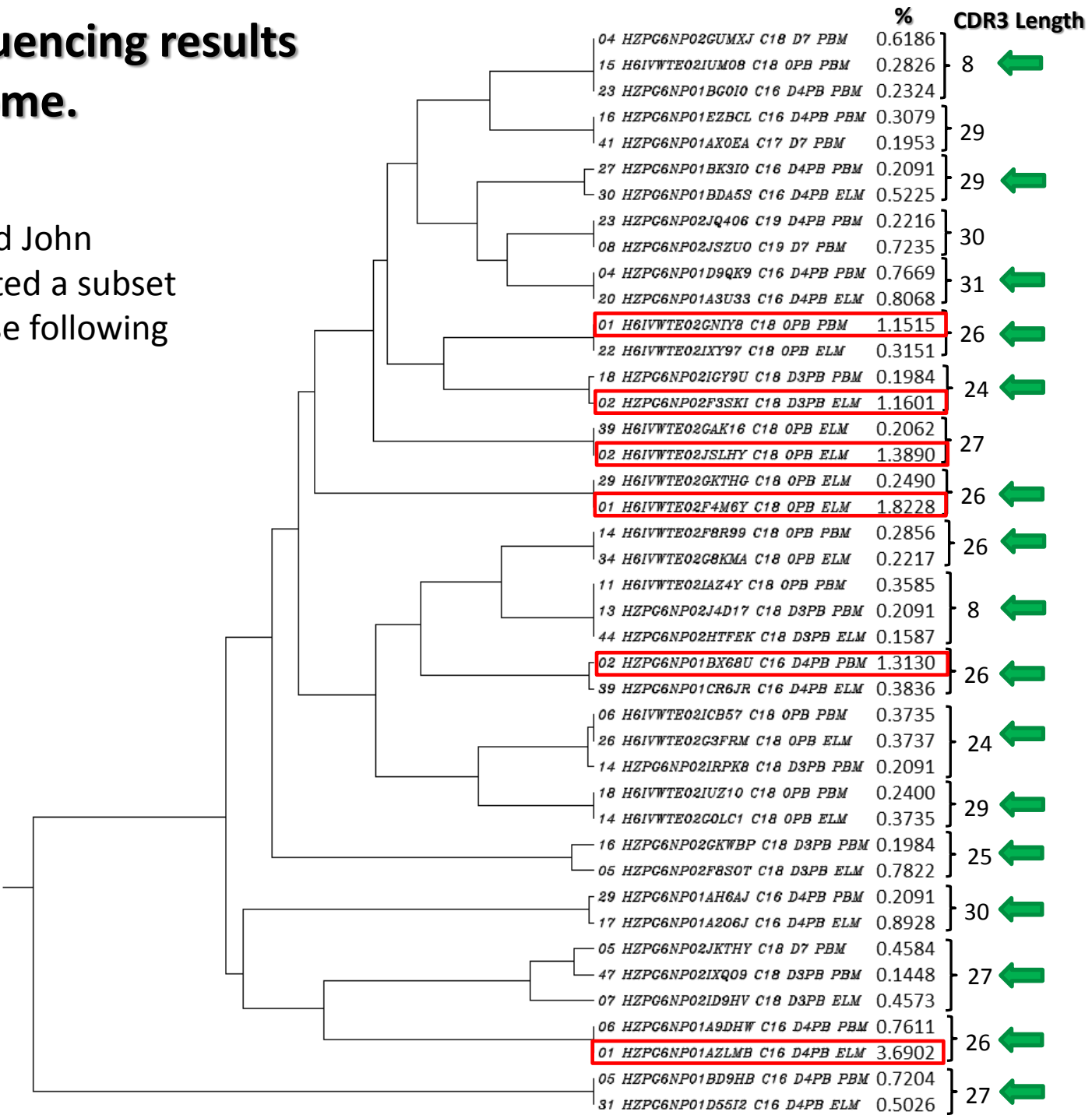


The small fraction of sequences with ultralong CHD3 completely disappears post immunisation

# Preliminary sequencing results

–lots more to come.

With Clare Grant and John Hammond we selected a subset that seem to increase following vaccination





# Selected Heavy Chain sequences for scale up

Sequences	CHR3 Length	% Abundance
(01) >04_HZPG6NP02GUMXJ_FMDC18_D7_PBMC	8	0.6186
(02) >44_HZPG6NP02HTFEK_FMDC18_D3PB_ELMC	8	0.6620
(03) >02_HZPG6NP02F3SKI_FMDC18_D3PB_ELMC	24	1.1601
(04) >26_H6IVWTE02G3FRM_FMDC18_OPB_ELMC	24	0.3737
(05) >05_HZPG6NP02F8SOT_FMDC18_D3PB_ELMC	25	0.7822
(06) >01_HZPG6NP01AZLMB_FMDC16_D4PB_ELMC	26	3.6902
(07) >01_H6IVWTE02GNIY8_FMDC18_OPB_PBMC	26	1.1515
(08) >34_H6IVWTE02G8KMA_FMDC18_OPB_ELMC	26	0.2218
(09) >01_H6IVWTE02F4M6Y_FMDC18_OPB_ELMC	26	1.8228
(10) >31_HZPG6NP01D55I2_FMDC16_D4PB_ELMC	27	0.5026
(11) >05_HZPG6NP02JKTHY_FMDC18_D7_PBMC	27	0.4584
(12) >07_HZPG6NP02ID9HV_FMDC18_D3PB_ELMC	27	0.4573
(13) >30_HZPG6NP01BDA5S_FMDC16_D4PB_ELMC	29	0.5225
(14) >14_H6IVWTE02G0LC1_FMDC18_OPB_ELMC	29	0.3735
(15) >17_HZPG6NP01A2O6J_FMDC16_D4PB_ELMC	30	0.8928
(16) >20_HZPG6NP01A3U33_FMDC16_D4PB_ELMC	31	0.8068

# Selection of Light Chain

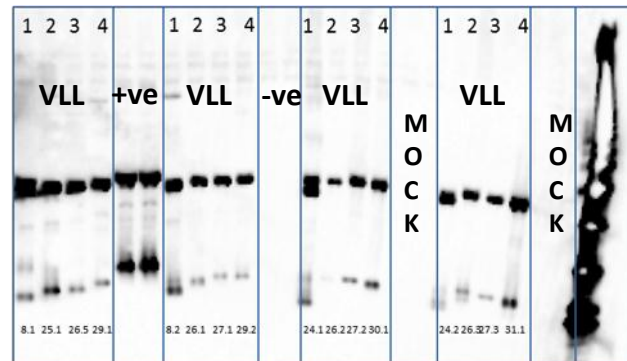
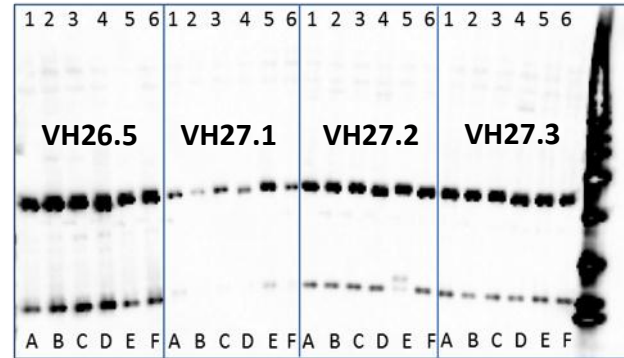
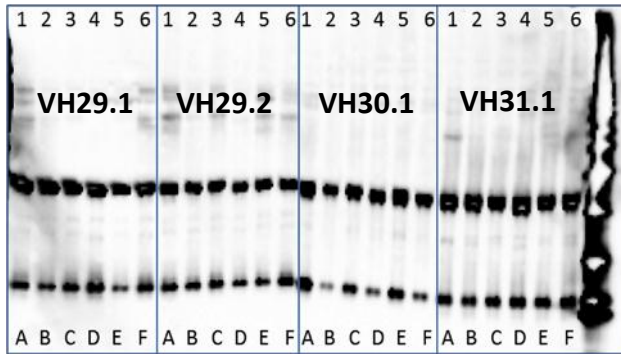
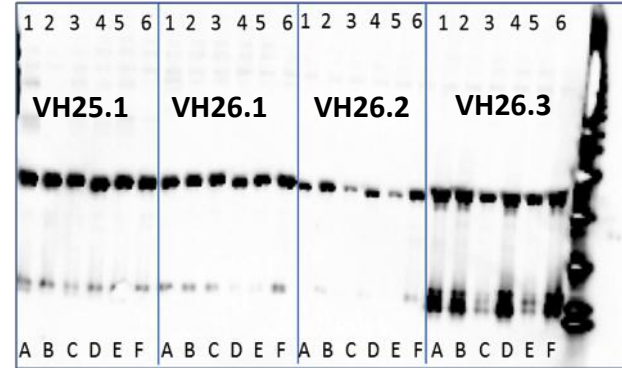
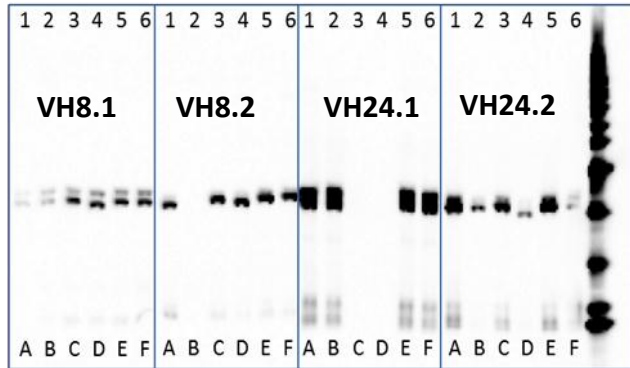
## Classified into 14 groups

```
A QAVLTQPSSVSGSLGQRVSITCSGSSSNVGTGNYVSWFQQIPGSAPRTLIYGATSRASGVPDRFSGSRSGNTATLTISLQAEDEADYFCASYQSG--NTAVFGSGTTLTVLGQPKSP-
B QAVLTQPSSVSGSLGQRVSITCSGSSNNIG-SYGVGWYQQVPGSGLRTIIYGSSSRPSGVPDRFSGSKSGNTATLTISLQAEDEADYFCATG DYSS-STAVFGSGTTLTVLGQPKSAP
C QAVLTQPSSVSGSLGQRVSITCSGSSSNVGNQ-YVSWYQLIPGSAPRTLIYGDTSRASGVPDRFSGSRSGNTATLTISLQAEDEADYFCASAEDSS-SNAVFGSGTTLTVLGQPKSPP
D QAVLTQPSSVSGSLGQRVSITCSGSSSNIG-SYNVGWYQQVPGSGLRTIIYGSSSRPSGVPDRFSGSKSGNTATLTISLQAEDEADYFCVAYDSSS-STAVFGSGTTLTVLGQPKSP-
E QAVLTQPSSVSGSLGQRVSITCSGSSSNVGYGNYVSWFQQIPGSAPRMLIYGATSRASGVPDRFSGSRSGNTATLTISLQAEDEADYFCASPDSSSS--GVFGSGTTLTVLGQPKSPP
F QAVLTQPSSVSGSLGQRVSITCSGSSNNIG-RYGVGWYQQVPGSGLRTIIYGSSSRPSGVPDRFSGSKSGNTATLTISLQAEDEADYFCAAGDSSS-STAVFGSGTTLTVLGQPKSPP
G QAVLTQPSSVSGSLGQRVSITCSGSSNNIG-SYGVGWYQQVPGSGLRTIIYGSSSRPSGVPDRFSGSKSGNTATLTISLQAEDEADYFCAAGDSSS-STAVFGSGTTLTVLGQPKSPP
H QAVLTQPSSVSGSLGQRVSITCSGSSSNIG-SYDVGWYQQVPGSGLRTIIYGSSSRPSGVPDRFSGSKSGNTATLTISLQAEDEADYFCAAGDSSS-STAVFGSGTTLTVLGQPKSPP
I QAVLTQPSSVSGSLGQRVSITCSGSSSNVGTGNYVSWFQQIPGSAPRTLIYGATSRASGVPDRFSGSRSGNTATLTISLQAEDEADYFCASYQSD--NTAVFGSGTTLTVLGQPKSAP
J QAVLTQPSSVSGSLGQRVSITCSGSSSNVGYGNYVSWFQQIPGSAPRMLIYGATSRASGVPDRFSGSRSGNTATLTISLQAEDEADYFCASPDSSSSGYAVFGSGTTLTVLGQPKSPP
K QAVLTQPSSVSGSLGQRVSITCSGSSSNVGRGNYVNWVQQIPGSAPRTLIYGATSRASGVPDRFSGSRSGNTATLTISLQAEDEADYFCAAYDSSS-NNAVFGSGTTLTVLGQPKSPP
L SYELTQPTSVSVALGQTAKITCSG---DLLDEQYQWYQQKPGQGPVVRVIYKDSERPSGISDRFSGSSSGKTATLTISGAQTEDEADYQCQADSSD--NAVFGSGTTLTVLGQPKSPP
M SYELTQPTSVSVALGQTAKITCSG---DLLDEQYQWYQQKPGQGPVVRVIYKDSERPSGISDRFSGSSSGKTATLTISGAQTEDEADYQCQADSSD--NPVFGSGTTLTVLGQPKSPP
N QAVLTQPSSVSGSLGQSVSITCSGSSSNVGNQ-YVSWYQMTIPGSAPRTLIYGDTSRASGVPDRFSGSRSGNTATLTISLQAEDEADYFCASAEDSS-SNAVFGSGTTLTVLGQPKSP-
. ****:* ** :** ..***** :: .*: * **.. :** :*.**:* ***** **.******. *.*****:* : : . **********.
```

## Wang *et al.* VL is closely related to group C

```
C QAVLTQPSSVSGSLGQRVSITCSGSSSNVGNQYVSWYQLIPGSAPRTLIYGDTSRASGVPDRFSGSRSGNTATLTISLQAEDEADYFCASAEDSSSNAVFGSGTTLTVLGQPKSPP
4K3E EAVLNQPSVSGSLGQRVSITCSGSSSNVGNQYVSWYQLIPGSAPRTLIYGDTSRASGVPDRFSGSRSGNTATLTISLQAEDEADYFCASAEDSSSNAVFGSGTTLTVLGQPKSPP
:***.*****
```

# Expression test of recombinant Fabs (all vs all)





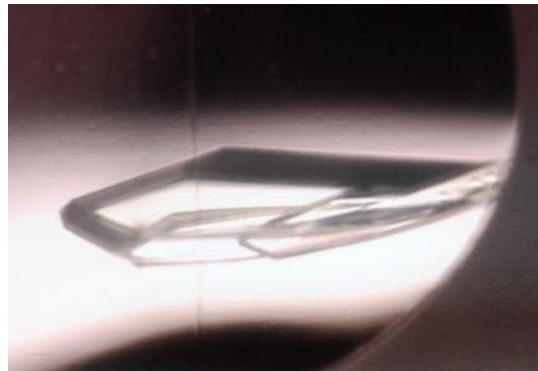
# Bovine Fab Crystals of selected Vh paired with Vl group E

**AK8.2\_VLE**



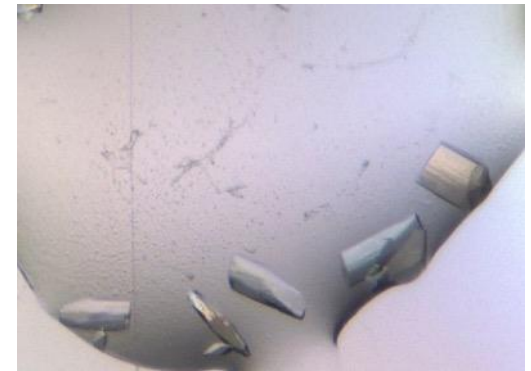
0.1M citric acid **pH 4.0**  
30 %w/v PEG 6000

**AK24.1\_VLE**



25 %w/v PEG 1500  
0.100 M SPG System **pH 5.0**

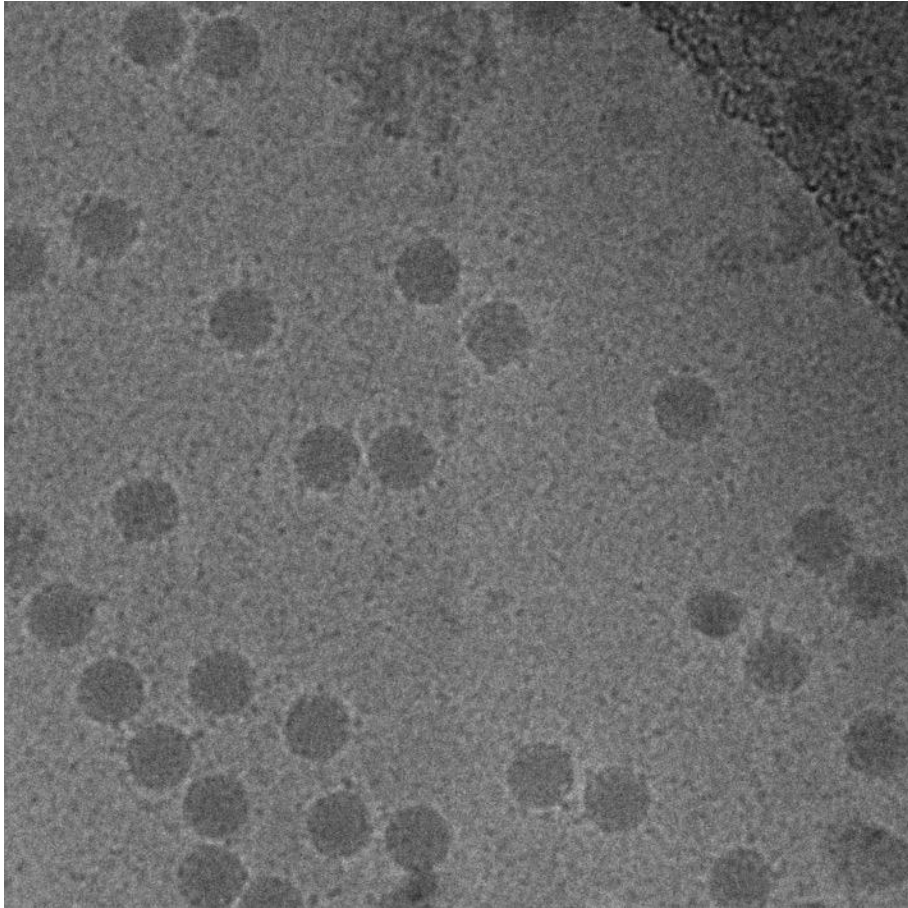
**AK26.1\_VLE**



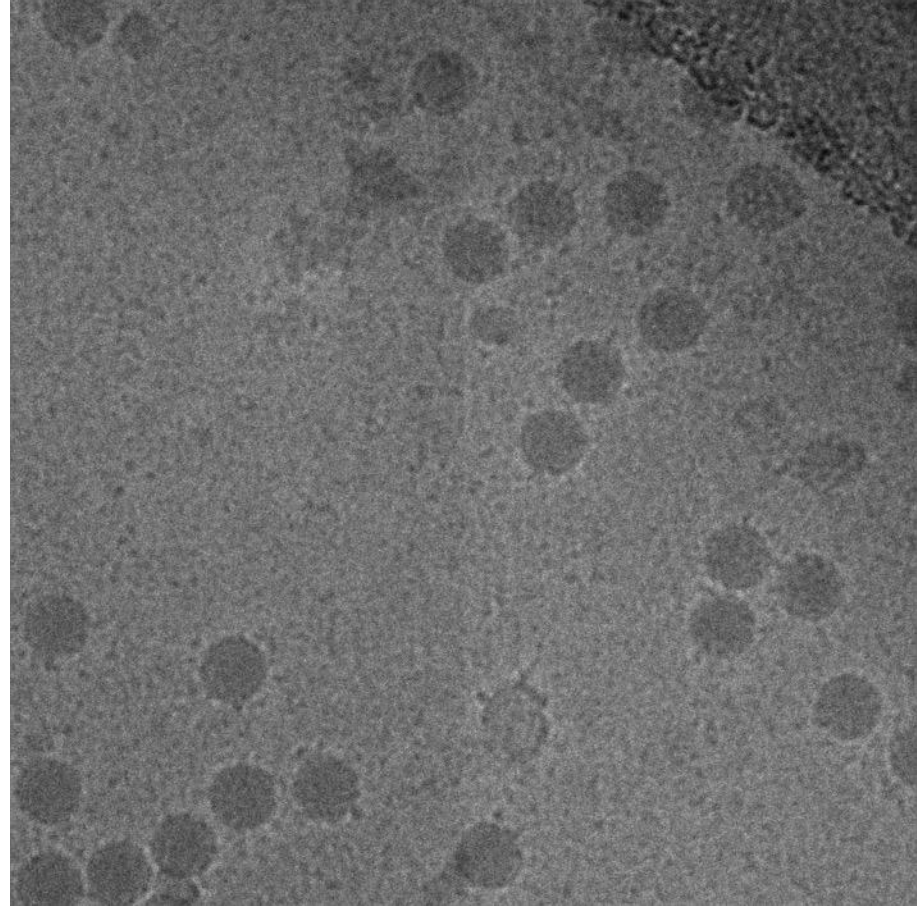
28.0% w/v PEG MME 2000  
0.1 M bis-Tris **pH 6.5**

Diffraction data measured at I03, I04 and I24 beamlines at the Diamond Light Source

# Fabs bind



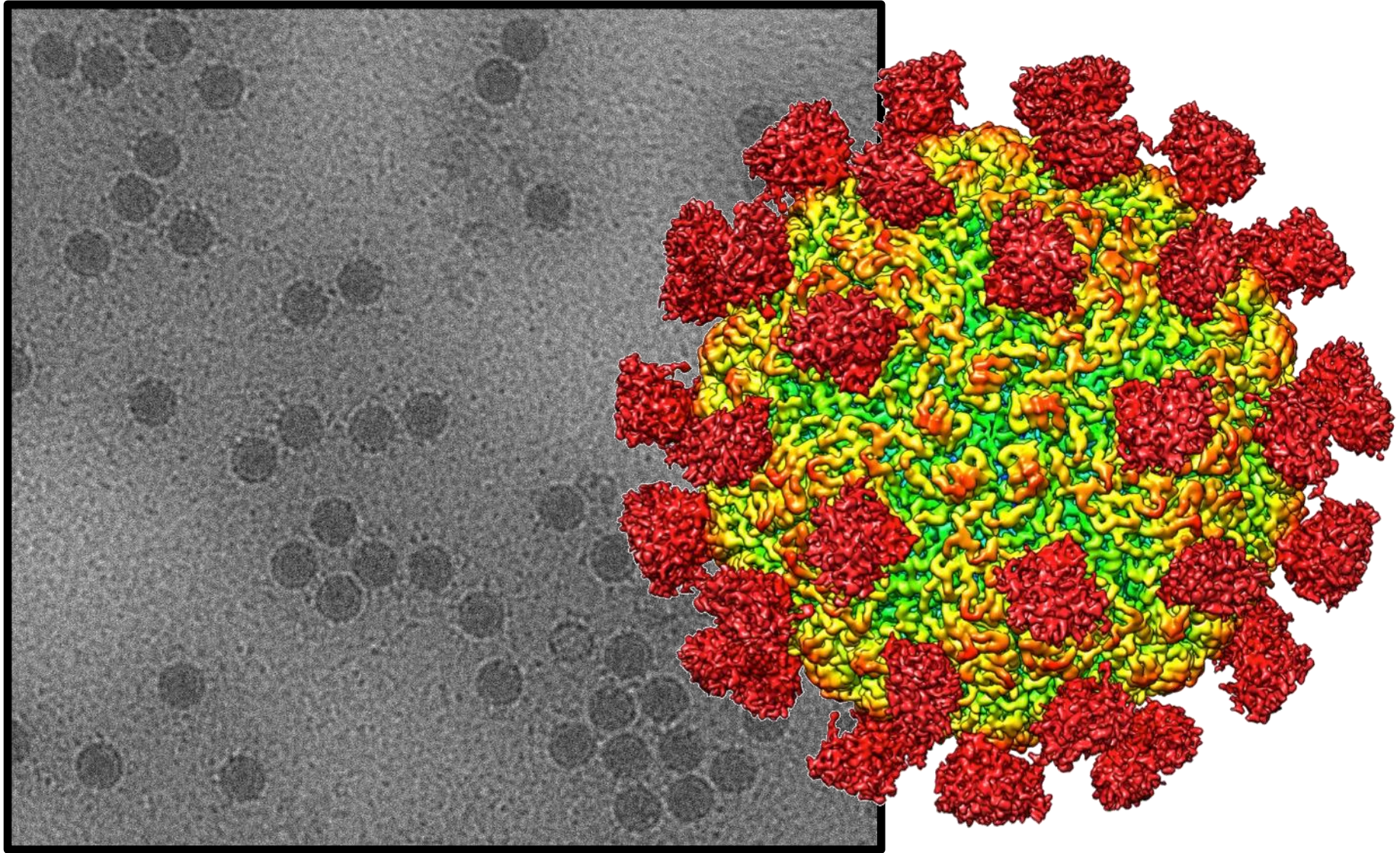
**O1M – IB11 complex**



**O1M – rFab(3) complex**



# 70hrs Collection on Polara



We now have EM data for 12 Fabs (CRD3 heavy chain loop form 8 to 31 residues) and apart from one, all are binding! - despite no careful light chain matching ... (structure determinations heating our building as I speak...)

So our aim to to investigate the nature of the epitopes for different serotypes and see if there is scope for either

- i) Ablating dominant serotype specific sites  
(eg GH loop)
- ii) Assembling a chimeric surface taking  
distinct sites from different serotypes  
stitching them together (*cf* Novartis)

Of course we have no idea if any of this will work!



So perhaps we are beginning to understand complex pathogens well enough to apply computational tools and chemistry to fight them more effectively.

But there will be plenty of challenges – not least how do we produce more cross-protective vaccines!

# Additional acknowledgements

## **Diamond:**

I24 team: Gwyndaf Evans, Danny Axford, David Waterman, James Foadi, Jun Aishima, Robin Owen

I03 team: Katherin McAuley, James Nicolson, Mark Williams (& from I04: Dave Hall)

Martin Walsh

**Strubi Oxford:** Liz Fry, Karl Harlos, Jon Grimes, Jingshan Ren, Claudine Porta, Abhay Kotcha

**Others:** OPPF-UK (Ray Owens *et al*, RC@H), MPL (Isabel de Moraes *et al*, Diamond/RC@H), groups from Leeds IMM, IAH Pirbright, Rao Zihe group, Beijing, Kay Grunewald, Helen Saibil....

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**Thank you**